Investigations on Leaving Group Based Intra- versus Intermolecular Glycoside Bond Formation

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Dedicated to Professor Richard Neidlein on the occasion of his 70th birthday

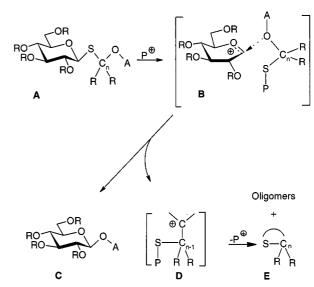
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Ligation of the glycosyl donor to the glycosyl acceptor through the leaving group was performed with the aim of enforcing glycoside bond formation by an intramolecular (1.x)-shift. To this end, syntheses of alkoxymethyl (2a and b), 2-alkoxyethyl (10a and b, 13a, 16a), 3-alkoxypropenyl (27a and b), and 7-alkoxy-4-oxaheptadienyl thioglucopyranoside derivatives (35a,b and d) were carried out. Their activation

with thiophilic promoter systems gave the expected glucopyranosides 5a, b and d in up to high chemical yields, yet modest anomeric control. Competition experiments showed that an intermolecular reaction course is favored in these reactions, although model considerations imply that convenient intramolecular (1,3)-, (1,4)-, (1,5)-, and (1,9)-shifts, respectively, of the glycosyl donor to the acceptor are possible.

Introduction

Glycosyl transfer within the active site of a glycosyltransferase can be regarded as an intramolecular process in which the anomeric center of the glycosyl donor and the accepting moiety are held in close proximity, thus enforcing the regio- and diastereoselectivity of the reaction.^[1,2] For corresponding in-vitro intramolecular glycosylations, linkers/spacers are required to connect the donor and the acceptor moieties appropriately. Linkers have been attached to functional groups on the donor; [3-15] mainly to the hydroxy group vicinal to the anomeric center, [3-10] thus preferentially furnishing either α - or β -products, depending on the configuration of the 2-hydroxy group. Particularly versatile and high yielding was the "rigid spacer" concept, recently introduced by us, which provides α - or β -products as desired.[15] We have also investigated linkage of the donor and the acceptor via the leaving group (leaving group based concept), which is rather attractive for preparative reasons.[11,13] However, success in this endeavor has thus far been erratic, because competition was occurring between intra- and intermolecular processes.^[13] Building on this concept, we have now carried out a study using as glycosyl donors different thioglycosides bearing the glycosyl acceptor by means of a C_n bridge (n = 1,2,3, and 7) at the sulfur atom. [16,17] As shown in Scheme 1, after generation of a glycosyl cation intermediate B from A, with the help of a thiophilic promoter P, intramolecular attack could take place at the accepting oxygen atom, yielding glycoside C by an (n + 2)-shift with release of a (stabilized) carbenium ion **D**, and this in turn could provide **E** and **P**. The results of our study are communicated in this paper.



Scheme 1. General reaction scheme

Results and Discussion

For the construction of starting materials with a C_1 bridge, 2,3,4,6-tetra-O-benzyl-1-thioglucose (1)^[18] (Scheme 2) was treated with chloromethyl ethyl ether in the presence of sodium hydride as base to afford ethoxymethyl thioglucoside 2a directly. The corresponding O,S-acetal 2b, with a sugar residue as acceptor, was obtained from methyl 6-O-acetyl-2,3,4-tri-O-benzylglucoside (3), which was converted with the Tebbe reagent^[19] into enol ether 4. Treatment of 4 and $I^{[18]}$ with $I^{[18]}$ with $I^{[18]}$ complex as catalyst gave 2b in 56% yield (1:1 mixture of diastereoisomers). Reaction of 2a with various promoter systems (Table 1) furnished ethyl glucopyranosides $I^{[20]}$ in up to 69% yield; the $I^{[20]}$ the $I^{[20]}$ ratios exhibited only low anomer selectivity, al-

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Scheme 2. Synthesis of starting materials $\mathbf{2a}$ and \mathbf{b} and their transformations

Table 1. Glycosylation results with 2a and b in dichloromethane

Temp.	Promoter	Product	Yield (%)	α,β
room temp.	TMSOTf (0.7 equiv.)	5a	69	2:1
-10 °C	TMSOTf (1.0 equiv.)	5a	39	2:3
room temp.	ZnCl ₂ ·OEt ₂ (2.0 equiv.)	5a	46	1:3
room temp.	DMTST (1.1 equiv.)	5b	27	1:1

though retention of configuration had been expected to be preferred. Similar results were obtained with 2b, affording known disaccharides $5b\alpha$, β . [21,25]

Therefore, we turned our attention to C₂-bridged systems in which the anomeric oxygen atom could be situated quite close conformationally to the anomeric center, via five connected atoms (Scheme 3). To this end, racemic methyl mandelate $(9)^{[22]}$ was O-alkylated with ethyl trifluoromethanesulfonate (R^1 -OTf, R^1 = Et) and also with the 6-O-trifluoromethanesulfonate of methyl 2,3,4-tri-O-benzyl-α-D-glucopyranoside, [23] affording compounds 8a and b. Reduction with lithium aluminum hydride in THF furnished diol derivatives 7a and b, which upon treatment with methanesulfonyl (mesyl) chloride in pyridine gave mesylates 6a and b in quantitative yields. Treatment of these with 2,3,4,6-tetra-O-benzyl-1-thio-β-D-glucose^[18] (1), in DME/DMF mixtures as solvent and in the presence of NaH as base, afforded βthioglycosides 10a and b as mixtures of two diastereoisomers; 10b was separated by chromatography $(\rightarrow 10bh)$ and 10bl). Treatment of 10a with dimethyl(methylthio)sulfonium triflate (DMTST) as promoter, in dichloromethane as solvent and under standard conditions, [24] afforded ethyl glycosides 5aa/\beta in up to 80\% yield with the expected preference for the β-anomer (Table 2). In acetonitrile as solvent, lower yields but slightly higher β-selectivities^[11b,25] were observed. With 10b, the two diastereoisomers 10bh and 10bl were investigated separately. In acetonitrile as solvent, yields and anomer selectivity in products 5b was very similar, while in dichloromethane and 1,2-dichloroethane as

Scheme 3. Synthesis of starting materials 10a and b and their transformations

Table 2. Glycosylation results with 10a and 10b and DMTST (1.1 equiv.) as promoter

	Solvent	Temp.	Product	Yield (%)	α,β
10a 10b	CH ₂ Cl ₂ CH ₂ Cl ₂ MeCN CH ₂ Cl ₂ CICH ₂ CH ₂ Cl MeCN	room temp. 0 °C room temp. room temp. 50 room temp.	- 5a 5b 5b	80 0 49 51/43 ^[a] 60 44/44 ^[a]	1:2 - 1:3 1.5:1/1.4:1 ^[a] 2:1 1:3/1:2 ^[a]

[[]a] Results from the two diastereoisomers.

solvent, even a preference for the formation of the α -anomers was observed.

Because of the conformational flexibility in the C2bridged systems 10a and b, a related but more rigid cyclic system was investigated. To this end, trans-2-bromo-1-hydroxyindane (11)^[22] and its homologue 14^[26] (Scheme 4) were transformed using 1-thioglucoside 1[18] into cis-substituted derivatives 12h,l and 15, respectively. Treatment of these with ethyl iodide in the presence of NaH as base and 15-crown-5 in DMF as solvent furnished the desired model compounds 13a and 16a. From 13a under standard glycosylation conditions, [24] yields of 5aα,β were up to 80% (Table 3); however, only in acetonitrile at room temp. or in dichloromethane at -20 °C as solvents was the expected preference for the β-anomer observed. Transformation of 16a into 5a exhibited similar behavior, but somewhat lower product yields were obtained than those observed for the transformation of 13a.

The low anomer selectivities with systems **2**, **10**, **13**, and **16** were reason to doubt the intramolecular reaction course in the glycosylations. Therefore, system **13** was investigated more carefully. Proper design of competition experiments can readily distinguish between intramolecular and intermolecular reaction courses.^[13,17] To this end, two different glycosyl donor and acceptor moieties, each of approximately similar reactivity, have to be ligated through the leaving group. Tetra-*O*-benzylglucose **17**^[27] and tetra-*O*-(3-methylbenzyl)glucose **18**^[13,17] should fulfil this donor requirement (Scheme 5). Similar acceptor properties were ex-

Scheme 4. Synthesis of 13a and 16a and their transformation into $5a\alpha_s\beta$

Table 3. Formation of glycosides 5aα,β from 13a and 16a

	Solvent	Temp.	Promoter	Yield (%)	α,β
13a 16a	CH_2Cl_2	room temp. -20 °C room temp. room temp. room temp. room temp.	DMTST (1.1 equiv.) DMTST (2.0 equiv.) MeOTf (1.2 equiv.) MeOTf (2.0 equiv.) DMTST (1.5 equiv.) MeOTf (1.2 equiv.)	80 24 78 70 47 60	1:1 1:3 1:1 1:1 1:3 1:1

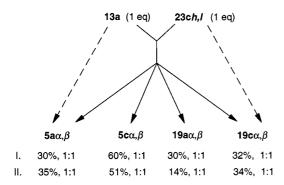
pected for primary alcohols ethanol and 2-bromobenzyl alcohol, which can readily be distinguished analytically. Transformation of 18 into trichloroacetimidate 20,[13,17] followed by reaction with ethanol under standard conditions, gave ethyl glucosides 19aα,β. The required Mbn-protected starting material was also obtained from 18; reaction with thionyl chloride, and then with potassium thioacetate, gave 1-acetylthio derivative 21. Removal of the S-acetyl group with sodium methoxide/methanol (similar to Zemplén conditions)[29] and immediate reaction with indane derivative 11 gave hydroxyindanyl thioglycosides 22h,l, which were separated by chromatography. Anomeric O-alkylation of **22h,** *l* with 2-bromobenzyl bromide furnished the desired starting material 23h,l required for the competition experiments. Activation of 23ch, l under standard conditions (DMTST, room temp., CH₂Cl₂) was also a means to obtain glycosides **19ca,** β (yield 64%, α : $\beta = 1:1$).

Competition experiments with 13a (1 equivalent) and 23ch, I (1 equivalent) in the presence of DMTST (total 5 equivalents) provided not only 5a and 19c, but also crossover products 5c and 19a (Scheme 6), thus clearly exhibiting that the glycosylations were mainly or exclusively following an intermolecular reaction course. In order to increase further the ring size for the potential intramolecular glycosyl transfer process, systems of structure 27 (Scheme 7), requiring a 1,5-shift, were investigated. This system, with the two heterocyclic rings in perpendicular arrangement, should also adopt a favorable conformation for the intramolecular glycosyl transfer; [17] facile cleavage of the C-O bond between the acceptor and the xanthene system should also be available, thus releasing the resonance-stabilized xanthylium moiety. For the synthesis of the starting material, 2-chloro-

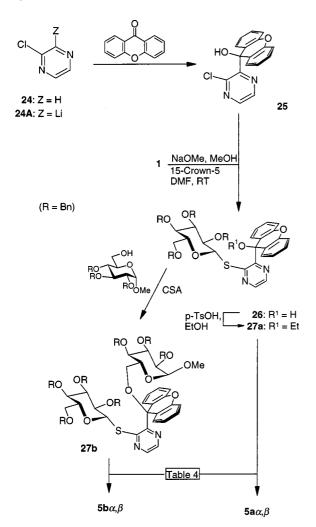
Scheme 5. Synthesis of 19a and c and 23c required for the competition experiments

23ch.l

pyrazine (24)^[22] was ortho-lithiated (\rightarrow 24A as intermediate);^[30] reaction with xanthenone furnished addition product 25 in 82% yield. Transformation of 1^[18] into the sodium salt, and dissolution in dry DMF led, with 25 as electrophile, to thioglycoside 26 (carbinol base). Acceptor attachment to the 10-position of the xanthenol moiety could readily be achieved by acid catalysis. Thus, with excess ethanol in the presence of pTsOH as catalyst, 10-Oethyl derivative 27a was obtained; similarly, treatment with methyl 2,3,4-tri-O-benzyl-α-D-glucopyranoside^[31] and camphorsulfonic acid (CSA) as catalyst in the presence of molecular sieves furnished the corresponding 10-O-(glucopyranosid-6-yl) derivative 27b. Transformation of 27a and 27b into glycosides 5a and 5b, respectively, could readily be accomplished with silver triflate as promoter (Table 4). Quite high product yields were obtained for both systems, but the α/β selectivities again were low. Addition of molecular sieves led to an increase of the retentive process, but these reactions were far from yielding a single product, as would



Scheme 6. Competition experiments with **13a** and 23*ch,l*; reagents and conditions: DMTST (5 equiv.), CH₂CI₂, room temp.; I: with **22ch**; II: with **22cl**



Scheme 7. Synthesis of 27a and b and their transformations

be expected for an ideal intramolecular glycosyl transfer to the acceptor.

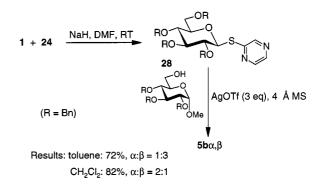
The pyrazinylthio group was investigated independently as a leaving group in glycosylation reactions (Scheme 8), since it should exhibit interesting properties.^[31]

To this end, 1-thioglucose derivative 1 was transformed with 24 into the desired glycosyl donor 28. In the context of this work, reaction with methyl 2,3,4-tri-O-benzyl- α -D-glucopyranoside^[32] as acceptor under the conditions de-

Table 4. Glycosylation results with 27a and 27b at room temperature; conc. ca. 5 \times $10^{-2}~\mbox{M}$

Solvent	Promoter ^[a]	Product	Yield [%]	α,β
Toluene CH ₂ Cl ₂ CH ₂ Cl ₂ Toluene Toluene	AgOTf (1+2 equiv.) AgOTf (3 equiv.) AgOTf (3 equiv.) MeOTf ^{[b][c]} (1.2 equiv.) AgOTf (3 equiv.) AgOTf (3 equiv.) AgOTf (3 equiv.)	5a 5a 5a 5a 5b 5b	64 74 82 10 66 69 75	1:3 1:3 3:2 1:1 1:7 1:3 3:2

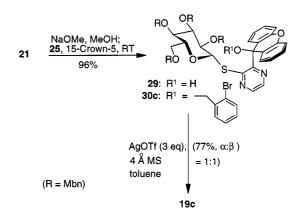
[a] With molecular sieves (4 Å). – [b]With 2,6-di-*tert*-butylpyridine (1.2 equiv.). – [c] Without molecular sieves.



Scheme 8. Thioglycoside 28 as glycosyl donor

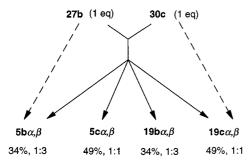
scribed in Table 4 was investigated. The results were very similar, thus supporting the intermolecular reaction course of the transformations of 27a and b.

This expectation was confirmed by competition experiments (Scheme 9). To this end, Mbn-protected 1-acetylthioglucose 21 was transformed into the sodium thiolate and then treated with 2-chloropyrazine derivative 25 to afford thioglycoside 29; reaction with 2-bromobenzyl alcohol in the presence of molecular sieves and CSA as catalyst furnished the desired starting material 30c for the competition experiments. In order to prove the efficiency of 30c as glycosyl donor/acceptor unit, activation with silver triflate was performed, leading to 19c in good yield. For reasons of comparison, disaccharide 19b^[13] was also required.



Scheme 9. Synthesis of 19c and 30c required for the competition experiments

With these compounds in hand, competition experiments could be carried out (Scheme 10). Treatment of **27b** and **30c** (one equivalent each) under standard glycosylation conditions furnished not only glycosides $5b\alpha$, β and $19c\alpha$, β but also the crossover products $5c\alpha$, β and $19b\alpha$, β in similar yields and anomer ratios. Once more, therefore, at least the prevalence of the intermolecular reaction course had been proven.



Scheme 10. Competition experiments with **27b** and **30c**; reagents and conditions: AgOTf(6 equiv.), molecular sieves (4 Å), toluene, room temp.

In none of the systems 2, 10, 13, and 27 – with C_1 , C_2 , and C₃ bridges, respectively – was the expected retention of configuration at the anomeric center observed in glycoside bond formation; obviously, intramolecular 1,3-, 1,4-, or 1,5-glycosyl cation shifts were disfavored. Therefore, systems capable of producing medium-sized rings in the glycosyl transfer were planned. In this way, S_N2-type shifts between donor and acceptor, as shown with transition state F in Scheme 11, might be accessible, leading to inversion of configuration at the anomeric center. To this end, 1,8-dicarboxyxanthene 31^[22] (Scheme 11) was reduced to diol 32, which upon treatment with thionyl chloride gave 1,8bis(chloromethyl) derivative 33. Compound 33 could be treated with methyl 2,3,4-[32] and 2,3,6-tri-O-benzyl-α-Dglucopyranoside, [33] to produce mainly monoethers 35b and d. Reaction of 32 with ethyl triflate in the presence of NaH as base afforded 34a, which, on treatment with thionyl chloride, gave chloromethyl derivative 35a, thus offering an alternative route for the synthesis of compounds of type 35. Reaction of 35a,b and d with 1[18] and NaH as base furnished target molecules 36a,b and d. Activation of these, again under standard glycosylation conditions, gave glycosides $5a\alpha,\beta,b\alpha,\beta$ and $d\alpha,\beta$ (Table 5). However, neither the epected preference for the inversion products (i.e., the α -anomers) nor even high product yields were observed. Instead, upon activation of, for instance, 36b with N-iodosuccinimide (NIS) in the presence of 2,6-di-tert-butyl-4-methylpyridine (DTP), a large amount (86%) of sulfenamide 37b was obtained, thus precluding the envisaged intramolecular reaction course. Possibly, the glycosyl donor moiety and the glycosyl acceptor moiety populate conformationally different sides of the flat tricyclic xanthene system (this is also supported by model calculations), hence once again disfavoring generation of transition state **F**, required for the intramolecular glycosyl transfer to the acceptor.

Scheme 11. Synthesis of 36a,b and d and their transformations

Table 5. Glycosylation results with 36a,b and d in dichloromethane

Temp.	Promoter	Product	Yield ([%)	α,β
0 °C	DMTST (1.4 equiv.) DMTST (1.2 equiv.) DMTST (2 equiv.) DMTST (1.5 equiv.)	5a	64 74 36 15	1:1 2:3 5:2 3:2

In conclusion, the leaving group based glycosylations studied in this paper follow an intermolecular rather than an intramolecular reaction course, although intramolecular (1,3)-, (1,4)-, and (1,5)-shifts (Schemes 2–4 and Scheme 7)

of the donor to the acceptor seem to be readily available. Activation of the glycosyl donor moiety in these systems obviously leads to solvent-and/or counterion-stabilized intermediates, which experience a lifetime long enough to search intermolecularly for sites of reactivity. This finally leads to product formation in up to high yields; yet the desired stereocontrol of an intramolecular reaction course is not attained. Of particular interest is the xanthene-derived system (Scheme 11) in which, according to model considerations, an intramolecular inversion 1,9-shift of the donor to the acceptor could take place, yet the results do not support this reaction course. Presumably, the 10-membered cyclic transition state as shown in F (Scheme 11) is conformationally disfavored. Obviously, structural modification of the xanthene skeleton could consider these conformational aspects, thus enforcing intramolecular leaving group based glycosylations. The xanthene system is also valuable for "rigid spacer based" intramolecular glycosylations, [15] which turned out to be very successful.

Experimental Section

General Methods: Optical rotations were determined with a Perkin–Elmer 241 MC polarimeter at 20° C. $^{-1}\mathrm{H}$ NMR spectra were recorded (internal Me₄Si) with a Bruker WM 250 Cryospec and a Bruker DRX 600 instrument. TLC was performed on silica gel 60 F₂₅₄ (Merck). – Chromatography was performed with silica gel (Baker, particle size 40 μm). Chromatography under elevated pressure (MPLC) was performed with LiChroprep Si 60 (Merck; 15–25 μm). The boiling range of the petroleum ether was 35–60 °C; anhydrous solvents were used. Molecular sieves (beads) were activated (250 °C; HV) before use; silver triflate was dried over KOH.

Ethoxymethyl 2,3,4,6-Tetra-O-benzyl-1-deoxy-1-thio-β-D-glucopyranoside (2a): To compound $\mathbf{1}^{[18]}$ (0.12 g, 0.22 mmol) in CH_2Cl_2 was added NEt₃ (45 µL, 0.32 mmol) and chloromethyl ethyl ether (30 μL, 0.32 mmol). The solution was diluted with ethyl acetate and washed with water. The organic layer was concentrated in vacuo. The pure product 2a (0.14 g, 100%) was isolated as a colorless oil. – TLC (petroleum ether/diethyl ether, 2:1): $R_f = 0.36$. – $[\alpha]_D =$ -24.7 (c = 1.0, CHCl₃). $- {}^{1}$ H NMR (250 MHz, CDCl₃): $\delta = 1.17$ $(t, {}^{3}J = 7.1 \text{ Hz}, 3 \text{ H}, \text{CH}_{3}), 3.49 - 3.57, 3.67 - 3.75 (2 \text{ m}, 8 \text{ H}, 2 - \text{H}, 3 \text{ H}, 3 + \text{H}, 3 +$ 3-H, 4-H, 5-H, 2 6-H, CH_2Me), 4.53 (d, $^2J = 12.2$ Hz, 1 H, SCH), $4.57 \text{ (d, } ^2J = 11.7 \text{ Hz}, 1 \text{ H, PhCH)}, 4.60 \text{ (d, } ^3J_{1,2} = 10 \text{ Hz}, 1 \text{ H, } 1-$ H), 4.61 (d, ${}^{2}J = 12.2 \text{ Hz}$, 1 H, SCH), 4.75 (d, ${}^{2}J = 11.7 \text{ Hz}$, 1 H, PhCH), 4.77 (d, ${}^{2}J = 10.4 \text{ Hz}$, 1 H, PhCH), 4.81 (d, ${}^{2}J = 11.0 \text{ Hz}$, 1 H, PhCH), 4.85 (d, ${}^{2}J$ = 11.0 Hz, 1 H, PhCH), 4.91 (d, ${}^{2}J$ = 11.0 Hz, 1 H, PhCH), 4.92 (d, ${}^{2}J$ = 10.3 Hz, 1 H, PhCH), 5.04 (d, $^{2}J = 11.5 \text{ Hz}, 1 \text{ H}, \text{ PhCH}, 7.15-7.37 (m, 20 \text{ H}, 4 \text{ Ph}).$ C₃₇H₄₂O₆S (614.8): calcd. C 72.28, H 6.89; found C 72.11, H 6.88.

Methyl 2,3,4-Tri-*O*-benzyl-6-*O*-[2-(2,3,4,6-tetra-*O*-benzyl-β-D-glucopyranosylthio)prop-2-yl]- α -D-glucopyranoside (2b): To compounds 4 (1.0 g, 1.98 mmol) and 1 (1.06 g, 1.90 mmol) in dioxane was added zinc chloride—diethyl ether (2.5 m in Et₂O, 0.2 mL, 0.5 mmol), while stirring. After 24 h, the mixture was neutralized with sat. NaHCO₃ solution. The mixture was diluted with ethyl acetate. The organic layer was separated; concentration in vacuo and then chromatography (petroleum ether/diethyl ether, 2:1) yielded 2b (1.17 g, 56%) as a slightly yellow oil. — TLC (petroleum ether/diethyl ether,

2:1): $R_{\rm f}=0.28.-{}^{1}{\rm H}$ NMR (250 MHz, CDCl₃): $\delta=1.48$ (s, 3 H, CH₃), 1.68 (s, 3 H, CH₃), 3.19 (s, 3 H, OCH₃), 3.32–3.72, 3.88–3.95 (2 m, 12 H, 2-H, 3-H, 4-H, 5-H, 2 6-H, 2'-H, 3'-H, 4'-H, 5'-H, 2 6'-H), 4.48–4.98 (m, 16 H, 1-H, 1'-H, 7 PhCH₂), 7.16–7.35 (m, 35 H, 7 Ph).

Methyl 6-*O*-Acetyl-2,3,4-tri-*O*-benzyl-α-D-glucopyranoside (3): To methyl 2,3,4-tri-*O*-benzyl-α-D-glucopyranoside^[32] (1.0 g, 2.15 mmol) in pyridine (10 mL) was added acetic anhydride (5 mL). At the end of the reaction, water and ethyl acetate were added. The layers were separated and concentrated in vacuo. Chromatography (toluene/ethyl acetate, 10:1) yielded 3 (1.0 g, 92%) as a colorless oil. TLC (toluene/ethyl acetate, 6:1): $R_{\rm f} = 0.40. - {}^{\rm 1}{\rm H}$ NMR (250 MHz, CDCl₃): δ = 2.02 (s, 3 H, OAc), 3.37 (s, 3 H, OCH₃), 3.48 (dd, ${}^{\rm 3}J_{4,5} = 10.0$ Hz, ${}^{\rm 3}J_{4,3} = 9.0$ Hz, 1 H, 4-H), 3.53 (dd, ${}^{\rm 3}J_{2,1} = 3.6$ Hz, ${}^{\rm 3}J_{2,3} = 9.6$ Hz, 1 H, 2-H), 3.81 (ddd, ${}^{\rm 3}J_{5,6} = 3.8$ Hz, ${}^{\rm 3}J_{5,4} = 10.1$ Hz, 1 H, 5-H), 4.01 (dd, ${}^{\rm 3}J = 9.3$ Hz, 1 H, 3-H), 4.20–4.31 (m, 2 H, 2 6-H), 4.53–5.03 (m, 7 H, 1-H, 3 PhCH₂), 7.24–7.38 (m, 15 H, 3 Ph). The NMR spectroscopic data correspond to those reported in ref.^[34]

Methyl 2,3,4-Tri-O-benzyl-6-O-(propen-2-yl)-α-D-glucopyranoside (4): To compound 3 (1.20 g, 2.37 mmol) was added a solution of dimethyldicyclopentadienyltitanium^[19] (1.0 g, 4.8 mmol in 10 mL toluene) and the mixture was warmed up to 70 °C. After 15 h, dimethyldicyclopentadienyltitanium (0.5 g, 2.4 mmol in 5 mL toluene) was once again added. After 5 h, the solution was concentrated in vacuo. Chromatography (petroleum ether/diethyl ether, 3:1) yielded 4 (1.10 g, 92%) as a slightly yellow oil. - TLC (petroleum ether/diethyl ether, 2:1): $R_f = 0.42$. – ¹H NMR (250 MHz, CDCl₃): $\delta = 1.82$ (s, 3 H, CH₃), 3.37 (s, 3 H, OCH₃), 3.55 (dd, $^{3}J_{2,1} = 3.6 \text{ Hz}, \, ^{3}J_{2,3} = 9.6 \text{ Hz}, \, 1 \text{ H}, \, 2\text{-H}), \, 2.64 \, (\text{dd}, \, ^{3}J_{4,3} = 9.1 \text{ Hz},$ ${}^{3}J_{4,5} = 9.1 \text{ Hz}, 1 \text{ H}, 4\text{-H}), 3.79 - 3.87 \text{ (m, 5 H, C=CH}_{2}, 5\text{-H}, 2 \text{ 6-}$ H), 3.66 (dd, ${}^{3}J_{3,2} = 9.1 \text{ Hz}$, ${}^{3}J_{4,5} = 9.1 \text{ Hz}$, 1 H, 3-H), 4.51 (d, $^{2}J = 10.7 \text{ Hz}, 1 \text{ H}, \text{ PhCH}), 4.61 (d, {}^{3}J_{1.2} = 3.6 \text{ Hz}, 1 \text{ H}, 1\text{-H}), 4.66$ $(d, {}^{2}J = 12.1 \text{ Hz}, 1 \text{ H}, \text{PhCH}), 4.78-4.86 \text{ (m, 3 H, 3 PhCH)}, 4.98$ $(d, {}^{2}J = 10.8 \text{ Hz}, 1 \text{ H}, \text{PhCH}), 7.22-7.38 \text{ (m, 15 H, 3 Ph)}.$

Ethyl 2,3,4,6-Tetra-O-benzyl- α/β -D-glucopyranoside (5 $\alpha\alpha,\beta$). — General procedure for Table 1: To compound 2a (0.16 g, 0.28 mmol) in CH₂Cl₂ (2 mL) was added DMTST (77 mg, 0.31 mmol). At the end of the reaction, sat. NaHCO₃ solution was added. The layers were separated and concentrated in vacuo. Chromatography (toluene/ ethyl acetate, 20:1) yielded $5a\alpha,\beta$ (86 mg, 69%, $\alpha:\beta = 1:2$) as a colorless oil. - General procedure for Table 2: To compound 10a (123 mg, 0.17 mmol) in CH₂Cl₂ (1 mL) was added DMTST (49 mg, 0.19 mmol in 1 mL CH₂Cl₂). After the end of the reaction, sat. NaHCO₃ solution was added. The layers were separated and concentrated in vacuo. Chromatography (toluene/ethyl acetate, 20:1) yielded $5a\alpha,\beta$ (77 mg, 80%, $\alpha:\beta = 1:1$) as a colorless oil. – General procedure for Table 3: To compound 13a (0.16 g, 0.28 mmol) in CH₂Cl₂ (2 mL) was added DMTST (77 mg, 0.31 mmol). After the end of the reaction, sat. NaHCO₃ solution was added. The layers were separated and concentrated in vacuo. Chromatography (toluene/ethyl acetate, 20:1) yielded $5a\alpha,\beta$ (86 mg, 69%, $\alpha/\beta = 1:2$) as a colorless oil. The physical data of $5\alpha\alpha,\beta$ are in accordance with literature data.[20]

Methyl 6-*O*-(2,3,4,6-Tetra-*O*-benzyl- α /β-D-glucopyranosyl)-2,3,4-tri-*O*-benzyl- α -D-glucopyranoside (5b α ,β): The title compound was prepared following the general procedure (Table 1 and 2) given above for 5a α ,β. The physical data are in accordance with literature data.^[21] — General procedure for Table 4: Molecular sieves (4 Å, ca. 0.7 g) and 27b (90 mg, 0.07 mmol) were stirred in CH₂Cl₂ (1.5 mL) for 15 min at room temp. After addition of silver triflate

(54 mg, 0.21 mmol), the reaction mixture was stirred for 24 h under exclusion of light. CH_2Cl_2 and sat. $Na_2S_2O_3$ solution were added to the dark orange reaction mixture, which was then stirred for 2 h. After the usual workup, chromatography (toluene/ethyl acetate, 12:1) yielded **5ba,β** (52 mg, 75%, α : β = 1.5:1).

From Compound 28: Methyl 2,3,4-tri-*O*-benzyl-α-D-glucopyrano-side^[31] (58 mg, 0.13 mmol), 28 (80 mg, 0.13 mmol), and molecular sieves (4 Å, ca. 0.5 g) were stirred in toluene (2.0 mL) for 30 min at room temp. After addition of silver triflate (90 mg, 0.35 mmol), the reaction mixture was stirred for 3 h under exclusion of light. Purification as described above yielded 5ba,β (90 mg, 72%, α:β = 1:3).

2-Bromobenzyl 2,3,4,6-Tetra-*O*-benzyl- α/β -D-glucopyranoside (5cα,β). – From Compounds 23c*l* and 13a: To compound 23c*l* (110 mg, 0.12 mmol) and compound 13a (86 mg, 0.12 mmol) in CH₂Cl₂ (2.0 mL) was added DMTST (0.25 m in CH₂Cl₂, 2.0 mL, 0.50 mmol) at room temp. After 45 min, sat. NaHCO₃ solution was added, followed by the usual workup. The resulting syrup was coevaporated twice with toluene. Chromatography with toluene/ethyl acetate, 60:1 first gave a fraction containing 19cα,β (31 mg, 34%, α : β = 1:1) and 5cα,β (43 mg, 51%, α : β = 1:1); changing to toluene/ethyl acetate, 30:1 gave a fraction containing 19aα,β (10 mg, 14%, α : β = 1:1) and 5aα,β (24 mg, 35%, α : β = 1:1).

From Compounds 23ch and 13a: To compound 23ch (110 mg, 0.12 mmol) and compound 13a (86 mg, 0.12 mmol) in CH₂Cl₂ (2.0 mL) was added DMTST (0.25 M in CH₂Cl₂, 2.0 mL, 0.50 mmol). After 45 min, the reaction mixture was purified as described above. Products were 19ca/ β (29 mg, 32%, α : β = 1:1), 5ca, β (51 mg, 60%, α : β = 1:1), 19aa, β (16 mg, 30%, α : β = 1:1), and 5aa, β (20 mg, 30%, α : β = 1:1).

From Compounds 27b and 30c: Molecular sieves (4 Å, 0.6 g), compound 30c (112 mg, 0.11 mmol), and compound 27b (135 mg, 0.11 mmol) were stirred in toluene (6 mL) for 10 min at room temp. After addition of silver triflate (163 mg, 0.63 mmol), the reaction mixture was stirred under exclusion of light. After 2 h, TLC indicated that 50% of the starting materials had been consumed. Also, however, two new, characteristically UV-active intermediates could be detected. Their $R_{\rm f}$ values fitted with those which would be expected for the transetherification products of 30c and 27b. After 24 h, CH₂Cl₂ (6 mL) and sat. Na₂S₂O₃ solution (8 mL) were added to the red-brown reaction mixture. After stirring for 4 h, the usual workup followed. Chromatography and analysis, as described earlier, yielded 19cα,β (40 mg, 49%, α/β = 1:1), 5cα,β (37 mg, 49%, α/β = 1:1), 19bα,β (38 mg, 34%), and 5bα,β (35 mg, 34%). A ratio of $\alpha/\beta = 1:3.5$ was determined for the last two.

Methyl *O*-(2,3,4,6-Tetra-*O*-benzyl- α /β-D-glucopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- α -D-glucopyranoside (5d α /β). — General procedure for Table 5: To compound 36d (90 mg, 0.07 mmol) in CH₂Cl₂ (2.0 mL) at room temp. was added DMTST (20 mg, 0.08 mmol). After 15 h, sat. NaHCO₃ solution was added, followed by the usual workup. Chromatography (toluene/ethyl acetate, 15:1) yielded first a fraction of 5d α ,β (10 mg, 15%, α :β = 1.5:1).

2-Ethoxy-2-phenylethyl Methanesulfonate (6a): To compound 7 (0.8 g, 4.8 mmol) in pyridine (8 mL) was added mesyl chloride (0.45 mL, 5.8 mmol). After 30 min, the solution was diluted with ethyl acetate. After filtration through silica gel, the solution was concentrated in vacuo. Chromatography (toluene/ethyl acetate, 20:1) yielded **7a** (1.18 g, 100%) as a colorless oil. – ¹H NMR (250 MHz, CDCl₃): δ = 1.20 (t, ${}^{3}J$ = 7.0 Hz, 3 H, CH₂CH₃), 3.00 (s, 3 H, SO₂CH₃), 3.37–3.55 (m, 2 H, CH₂CH₃), 4.22 (dd, ${}^{3}J$ =

3.8, ${}^{2}J = 11.2 \text{ Hz}$, 1 H, CHOMes), 4.33 (dd, ${}^{3}J = 8.1$, ${}^{2}J = 11.2 \text{ Hz}$, 1 H, CHOMes), 4.60 (dd, ${}^{3}J = 3.7 \text{ Hz}$, 1 H, PhCH), 7.15–7.42 (m, 5 H, Ph).

2-(Methyl 2,3,4-Tri-O-benzyl-α-D-glucopyranosid-6-yl)-2-phenylethyl Methanesulfonate (6bh,l): Compound 6bh was prepared using 7bh, following the procedure given for 6a; colorless solid, 67% yield. – TLC (toluene/ethyl acetate, 10:1): $R_f = 0.38$. – $[\alpha]_D =$ -6.0 (c = 1.0, CHCl₃). $- {}^{1}$ H NMR (250 MHz, CDCl₃): $\delta = 2.89$ (s, 3 H, SO₂CH₃), 3.36 (s, 3 H, OCH₃), 3.45-3.53 (m, 3 H, 2-H, 4-H, 6-H), 3.63 (dd, ${}^{3}J_{6,5} = 1.6 \text{ Hz}$, ${}^{2}J_{6,6'} = 11.1 \text{ Hz}$, 1 H, 6-H), 3.72 (ddd, ${}^{3}J_{5,6} = 1.6 \text{ Hz}$, ${}^{3}J_{5,6'} = 4.9$, ${}^{3}J_{5,4} = 10.2 \text{ Hz}$, 1 H, 5-H), 3.98 (dd, $J_{3,2} = 9.2 \text{ Hz}$, $J_{3,4} = 9.2 \text{ Hz}$, 1 H, 3-H), 4.21 (dd, ${}^{3}J =$ 4.8 Hz, ${}^{2}J = 11.0$ Hz, 1 H, CHOMes), 4.31 (dd, ${}^{3}J = 7.8$ Hz, ${}^{2}J =$ 11.0 Hz, 1 H, CHOMes), 4.54-4.61 (m, 3 H, 1-H, 2 PhCH), 4.66 $(d, {}^{2}J = 12.2 \text{ Hz}, 1 \text{ H}, PhCH), 4.79 (d, {}^{2}J = 12.2 \text{ Hz}, 1 \text{ H}, PhCH),$ 4.80 (d, ${}^{2}J = 10.9 \text{ Hz}$, 1 H, PhCH), 4.87 (d, ${}^{2}J = 11.1 \text{ Hz}$, 1 H, PhCH), 4.97 (d, ${}^{2}J = 10.9 \text{ Hz}$, 1 H, PhCH), 7.16–7.40 (m, 20 H, 4 Ph). - C₃₇H₄₂O₉S (662.8): calcd. C 67.05, H 6.39; found C 67.26, H 6.37.

Compound 6bl: prepared using **7bl**, following the procedure given for **6a**; colorless solid, 93% yield. – TLC (toluene/ethyl acetate, 3:2): $R_{\rm f}=0.60.$ – ¹H NMR (250 MHz, CDCl₃): δ 2.90 (s, 3 H, SO₂CH₃), 3.36 (s, 3 H, OCH₃), 3.47–3.99 (m, 6 H, 2-H, 3-H, 4-H, 5-H, 2 6-H), 4.21 (dd, ${}^3J=3.7$ Hz, ${}^2J=11.2$ Hz, 1 H, CHOMes), 4.37 (dd, ${}^3J=8.1$ Hz, ${}^2J=11.2$ Hz, 1 H, CHOMes), 4.42 (d, ${}^2J=10.8$ Hz, 1 H, PhCH), 4.57 (d, ${}^3J_{1,2}=3.5$ Hz, 1 H, 1-H), 4.66 (dd, ${}^3J=3.7$ Hz, ${}^3J=8.1$ Hz, 1 H, PhCHCH₂), 4.68 (d, ${}^2J=12.1$ Hz, 1 H, PhCH), 4.79 (d, ${}^2J=9.9$ Hz, 1 H, PhCH), 4.80 (d, ${}^2J=12.2$ Hz, 1 H, PhCH), 4.81 (d, ${}^2J=11.1$ Hz, 1 H, PhCH), 4.98 (d, ${}^2J=9.8$ Hz, 1 H, PhCH), 7.04–7.37 (m, 20 H, 4 Ph). – C₃₇H₄₂O₉S (662.8): calcd. C 67.05, H 6.39; found C 67.06, H 6.53.

2-Ethoxy-2-phenylethanol (7a): To a suspension of lithium aluminum hydride (0.18 g, 4.7 mmol) in dry Et₂O (10 mL) was added **8a** (1.3 g, 6.7 mmol) in Et₂O (7 mL). The mixture was refluxed. At the end of the reaction, ethyl acetate and water were added. The solid residue was dissolved in dilute sulfuric acid. The organic layer was separated and concentrated in vacuo. Chromatography (toluene/ ethyl acetate, 20:1) yielded **7a** (0.90 g, 81%) as a colorless oil. – TLC (toluene/ethyl acetate, 10:1). – $R_{\rm f}=0.23.$ – ¹H NMR (250 MHz, CDCl₃): δ 1.22 (t, ${}^3J=7.0$ Hz, 3 H, CH₃), 2.50 (br. s, 1 H, OH), 3.40 (dq, ${}^3J=7.0$ Hz, ${}^2J=9.3$ Hz, 1 H, OCHCH₃), 3.50 (dq, ${}^3J=7.0$ Hz, ${}^2J=9.3$ Hz, 1 H, OCHCH₃), 3.63–3.71 (m, 2 H, CH₂OH), 4.20 (dd, 1 H, PhCH, ${}^3J=4.3$ Hz, ${}^3J=8.1$ Hz). This compound had been prepared previously. [35] NMR-spectroscopic data had not been provided.

Methyl 2,3,4-Tri-O-benzyl-6-(2-hydroxy-1-phenylethyl)-α-D-glucopyranoside (7bh,l): The title compound was prepared from 8b, following the procedure described above for 7a, in 56% yield.

Compound 7bh: Colorless oil, TLC (toluene/ethyl acetate, 3:2): $R_{\rm f} = 0.38$. $- [\alpha]_{\rm D} = -19.5$ (c = 1.0, CHCl₃). $- {}^{1}{\rm H}$ NMR (250 MHz, CDCl₃): $\delta = 2.44$ (br. s, 1 H, OH), 3.37 (s, 3 H, OCH₃), 3.40–4.00 (m, 8 H, 2-H, 3-H, 4-H, 5-H, 2 6 H, CH₂OH), 4.38 (dd, ${}^{3}{\it J} = 3.9$ Hz, ${}^{3}{\it J} = 8.3$ Hz, 1 H, PhCHCH₂), 4.56 (d, ${}^{3}{\it J} = 11.0$ Hz, 1 H, PhCH), 4.61 (d, ${}^{3}{\it J}_{1,2} = 3.5$ Hz, 1 H, 1-H), 4.66 (d, ${}^{2}{\it J} = 12.2$ Hz, 1 H, PhCH), 4.68 (d, ${}^{2}{\it J} = 12.2$ Hz, 1 H, PhCH), 4.81 (d, ${}^{2}{\it J} = 10.8$ Hz, 1 H, PhCH), 4.88 (d, ${}^{2}{\it J} = 11.0$ Hz, 1 H, PhCH), 4.98 (d, ${}^{2}{\it J} = 10.8$ Hz, 1 H, PhCH), 7.16–7.37 (m, 20 H, 4 Ph). - C₃₆H₄₀O₇ (584.7): calcd. C 73.95, H 6.90; found C 73.51, H 6.87.

Compound 7b/: Colorless oil. – TLC (toluene/ethyl acetate, 3:2): $R_{\rm f} = 0.23. - [\alpha]_{\rm D} = 46.0 \ (c = 1.0, {\rm CHCl_3}). - {\rm ^1H} \ {\rm NMR} \ (250 \ {\rm MHz},$

CDCl₃): $\delta = 2.59$ (br. s, 1 H, OH), 3.39 (s, 3 H, OCH₃), 3.55 (dd, $J_{2,1} = 3.5$ Hz, $J_{2,3} = 9.6$ Hz, 1 H, 2-H), 3.61–3.99 (m. 7 H, 3-H, 4-H, 5-H, 2 6-H, C H_2 OH), 4.42–4.49 (m, 1 H, PhCHCH₂), 4.65 (d, ${}^3J_{1,2} = 3.3$ Hz, 1 H, 1-H), 4.68 (d, ${}^2J = 11.8$ Hz, 1 H, PhCH), 4.78–4.86 (m, 3 H, 3 PhCH), 4.99 (d, ${}^2J = 10.8$ Hz, 1 H, PhCH), 7.06–7.38 (m, 20 H, 4 Ph). – $C_{36}H_{40}O_7$ (584.7): calcd. C 73.95, H 6.90; found C 73.55, H 7.03.

Methyl 2-Ethoxy-2-phenylacetate (8a): To compound 9 (1.5 g, 9.0 mmol) in CH₂Cl₂ (50 mL) was added NaH (0.22 g, 9.0 mmol). At the end of H₂ evolution, ethyl triflate (1.42 mL, 9.9 mmol) was added. When the reaction was complete, sat. NH₄Cl solution was added. After extraction with ethyl acetate, the organic layer was concentrated in vacuo. Chromatography (toluene/ethyl acetate, 10:1) yielded 8a (1.35 g, 77%) as a colorless oil. – TLC (toluene/ethyl acetate, 10:1): $R_{\rm f} = 0.61$. – ¹H NMR (250 MHz, CDCl₃): $\delta = 1.28$ (t, $^3J = 7.0$ Hz, 3 H, CH₃), 3.59 (dq, $^3J = 7.0$ Hz, $^2J = 9.1$ Hz, 1 H, OCHMe), 3.61 (dq, $^3J = 7.0$, $^2J = 9.1$ Hz, 1 H, OCHMe), 3.71 (s, 3 H, OCH₃), 4.89 (s, 1 H, PhCH), 7.32–7.48 (m, 5 H, Ph). This compound had been prepared previously. [35] NMR-spectroscopic data had not been provided.

Methyl 2-(Methyl-2,3,4-tri-*O***-benzyl-α-D-glucopyranosid-6-yl)-2-phenylacetate (8b):** This compound was prepared using methyl 2,3,4-tri-*O*-benzyl-6-*O*-trifluoromethanesulfonyl-α-D-glucopyranoside^[23] and following the procedure given for **8a**; colorless oil, 76% yield. – TLC (toluene/ethyl acetate, 10:1): $R_{\rm f} = 0.32$. – $[\alpha]_{\rm D} = 20.0$ (c = 1.0, CHCl₃). – ¹H NMR (250 MHz, CDCl₃): δ = 3.35 (s, 3 H, OCH₃), 3.48–3.92 (m, 6 H, 2-H, 3-H, 4-H, 5-H, 2 6-H), 3.63, 3.64 (2 s, 3 H, CO₂CH₃), 3.99 (dd, $^{3}J = 9.2$ Hz, 1 H), 4.49–5.00 (m, 8 H, 1-H, 7 PhCH), 7.15–7.44 (m, 20 H, 4 Ph). – $C_{37}H_{40}O_{8}$ (612.7): calcd. C 72.53, H 6.58; found C 72.60, H 6.65.

2-Ethoxy-2-phenyl-1-(2,3,4,6-tetra-*O***-benzyl-1-deoxy-**β**-D-glucopyranosylthio)ethane (10ah,l):** To compound **1** (1.0 g, 1.8 mmol) in DME and DMF (30 mL, ratio 2:1) was added NaH (43 mg, 1.8 mmol). To the solution was added 15-crown-5 (0.36 mL, 1.8 mmol), and it was then warmed up to 60 °C. To the solution was added **6a** (0.5 g, 2.0 mmol). After 3 h, the solution was neutralized with sat. NH₄Cl solution. After extraction with ethyl acetate, the organic layer was concentrated in vacuo. Chromatography (toluene/ethyl acetate, 20:1) yielded **10a** (1.08 g, 85%).

Compound 10a*h*: Colorless oil. – TLC (toluene/ethyl acetate, 10:1): $R_{\rm f} = 0.34$. – $[\alpha]_{\rm D} = 6.0$ (c = 1.0, CHCl₃). $^{-1}{\rm H}$ NMR (250 MHz, CDCl₃): $\delta = 1.17$ (t, $^{3}J = 7.0$ Hz, 3 H, CH₃), 2.83 (dd, $^{2}J = 13.6$ Hz, $^{3}J = 4.8$ Hz, 1 H, SCH), 3.18 (dd, $^{2}J = 13.6$ Hz, $^{3}J = 8.3$ Hz, 1 H, SCH), 3.37 (q, $^{3}J = 7.0$ Hz, 2 H, OC $H_{2}{\rm Me}$), 3.41–3.47 (m, 2 H, 2-H, 5-H), 3.57–3.78 (m, 4 H, 3-H, 4-H, 2 6-H), 4.49–4.95 (m, 10 H, 1-H, 9 PhCH), 7.15–7.39 (m, 25 H, 5 Ph). – $C_{44}H_{48}O_{6}{\rm S}$ (704.9): C 74.97, H 6.86; found C 74.58, H 6.87.

Compound 10al: Colorless oil. — TLC (toluene/ethyl acetate): $R_{\rm f}=0.29.$ — [α]_D = -17 (c=1.0, CHCl₃). — ¹H NMR (250 MHz, CDCl₃): δ = 1.16 (t, $^3J=7.0$ Hz, 3 H, CH₃), 2.98 (dd, $^2J=11.5$ Hz, $^3J=6.1$ Hz, 1 H, SCH), 3.05 (dd, $^2J=11.5$ Hz, $^3J=6.1$ Hz, 1 H, SCH), 3.33—3.44, 3.59—3.72 (2 m, 8 H, CH₂Me, 2-H, 3-H, 4-H, 5-H, 2 6-H), 4.30 (d, $^3J_{1,2}=9.7$ Hz, 1 H, 1-H), 4.47—4.63 (m, 4 H, 4 Ph), 4.69 (d, $^2J=10.2$ Hz, 1 H, PhCH), 4.80 (d, $^2J=10.8$ Hz, 1 H, PhCH), 4.82 (d, $^2J=11.0$ Hz, 1 H, PhCH), 4.88 (d, $^2J=10.3$ Hz, 1 H, PhCH), 4.90 (d, $^2J=11.0$ Hz, 1 H, PhCH), 7.14—7.37 (m, 25 H, 5 Ph). — $C_{44}H_{48}O_6S$ (704.9): calcd. C 74.97, H 6.86; found C 74.71, H 6.94.

Methyl 6-O-[2-(2,3,4,6-Tetra-O-benzyl-1-deoxy-β-D-glucopyranosyl-thio)-1-phenylethyl]-2,3,4-tri-O-benzyl-α-d-glucopyranoside (10bh,l):

Compound **10bh** was prepared using **6bh**, following the procedure given for **10ah,I**; colorless oil, 36% yield. — TLC (toluene/ethyl acetate, 10:1): $R_{\rm f}=0.43.$ — ¹H NMR (250 MHz, CDCl₃): $\delta=2.99$ (dd, ${}^3J=6.0$ Hz, ${}^2J=13.4$ Hz, 1 H, SCH), 3.09 (dd, ${}^3J=7.3$ Hz, ${}^2J=13.4$ Hz, 1 H, SCH), 3.25 (dd, ${}^3J_{5,6}=2.6$ Hz, ${}^3J_{5,4}=9.2$ Hz, 1 H, 5a-H or 5b-H), 3.32 (s, 3 H, OCH₃), 3.36—3.65 (m, 10 H, 2a-H, 2b-H, 3a-H, 3b-H, 4a-H, 4b-H, 2 6a-H, 2 6b-H), 3.96 (ddd, ${}^3J_{5,6}=2.7$ Hz, ${}^3J_{5,4}=9.0$ Hz, 1 H, 5b-H or 5a-H), 4.16 (d, ${}^3J_{1b,2b}=9.6$ Hz, 1 H, 1b-H), 4.45—4.98 (m, 16 H, 1a-H, 15 PhCH), 7.12—7.23 (m, 40 H, 8 Ph). — $C_{70}H_{74}O_{11}S$ (1123.4): calcd. C 74.84, H 6.64; found C 74.35, H 6.62.

Compound 10bl: Prepared using **6bl**, following the procedure given for **10a**; colorless oil, 46% yield. – TLC (toluene/ethyl acetate, 10:1): $R_{\rm f}=0.43.-[\alpha]_{\rm D}=12.0$ (c=1.0, CHCl₃). – $^{1}{\rm H}$ NMR (250 MHz, CDCl₃): $\delta=2.88$ (dd, $^{3}J=5.4$ Hz, $^{2}J=13.3$ Hz, 1 H, SCH), 3.27 (dd, $^{3}J=7.7$ Hz, $^{2}J=13.3$ Hz, 1 H, SCH), 3.35 (s, 3 H, OCH₃), 3.40–3.76 (m, 3 H), 3.96 (dd, $^{3}J=9.3$ Hz, 1 H, 2a-H, 2b-H, 3a-H, 3b-H, 4a-H, 4b-H, 5a-H, 5b-H, 2 6a-H, 2 6b-H), 4.33 (d, $^{3}J=10.8$ Hz, 1 H), 4.49–4.97 (m, 16 H, 1a-H, 1b-H, 14 PhCH), 6.99–7.37 (m, 40 H, 8 Ph). – $C_{70}H_{74}O_{11}S$ (1123.4): calcd. C 74.84, H 6.64; found C 75.05, H 6.79.

D,L-erythro-1-Hydroxy-2-(2,3,4,6-tetra-O-benzyl-1-deoxy-β-D-glucopyranosylthio)indane (12h,l): To compound 1^[18] (0.18 g, 0.32 mmol) and 15-crown-5 (64 μL, 0.32 mmol) in DMF (2 mL) was added NaH (7 mg, 31 mmol). After 10 min, 2-bromo-1-hydroxyindane (11)^[22] (0.14 g, 0.65 mmol) was added; 10 min later, sat. NaHCO₃ solution was added. After extraction with ethyl acetate, the organic layer was concentrated in vacuo. Chromatography (petroleum ether/ethyl acetate, 4:1) yielded 12h,l (0.97 g, 78%) as a colorless oil. – TLC (toluene/ethyl acetate, 9:1): R_f = 0.38. – C₄₃H₄₄O₆S (688.9): calcd. C 74.97, H 6.44; found C 74.82, H 6.46. – The diastereomers were separated by MPLC (toluene/ethyl acetate, 20:1) and acetylated, to afford 12h-hc and 12l-hc, respectively.

Compound 12h: ¹H NMR (250 MHz, CDCl₃): $\delta = 3.03$ (dd, ²J = 15.6 Hz, ³J = 8.7 Hz, 1 H, SCHCH), 3.22 (dd, ²J = 15.8 Hz, ³J = 7.7 Hz, 1 H, SCH-CH), 3.39-3.92 (m, 7 H, SCH, 2-H, 3-H, 4-H, 5-H, 2 6-H), 4.49-5.05 (m, 10 H, 1-H, 9 PhCH), 7.12-7.40 (m, 24 H, C₆H₄, Ph).

Compound 12*h*-*Ac*: ¹H NMR (250 MHz, CDCl₃): δ = 2.00 (s, 3 H, Ac), 3.19–3.34 (m, 2 H, 2 3'-H), 3.40–3.77 (m, 6 H, 2-H, 3-H, 4-H, 5-H, 2 6-H), 3.86–3.95 (m, 1 H, SCH), 4.52–4.64 (m, 4 H, 1-H, 3 PhCH), 4.74–4.95 (m, 5 H, 5 PhCH), 6.20 (d, ³*J* = 5.6 Hz, 1 H, 1'-H), 7.16–7.49 (m, 24 H, C₆H₄, 4 Ph).

Compound 12: ¹H NMR (250 MHz, CDCl₃): δ = 2.99 (ddd, $J_{5,4}$ = 9.4 Hz, $J_{5,6}$ = 3.0 Hz, 1 H, 5-H), 3.13 (dd, ${}^{2}J$ = 16.5 Hz, ${}^{3}J$ = 5.3 Hz, 1 H, 3'-H), 3.34 (dd, ${}^{2}J$ = 16.6 Hz, ${}^{3}J$ = 7.7 Hz, 1 H, 3'-H), 2.26 (dd, ${}^{3}J$ = 9.6 Hz, 1 H), 3.55 (dd, ${}^{2}J$ = 9.0 Hz, 1 H), 3.61–3.68 (m, 5 H, 2-H, 3-H, 4-H, 2 6-H), 3.91 (ddd, ${}^{3}J$ = 7.8, 5.6 Hz, 1 H, SCH), 4.31 (d, ${}^{3}J_{1,2}$ = 9.8 Hz, 1 H, 1-H), 4.49–4.90 (m, 8 H, 8 PhCH), 5.18 (dd, ${}^{3}J$ = 6.7 Hz, 1 H, PhCHOH), 7.11–7.36 (m, 24 H, C₆H₄, 4 Ph).

Compound 12*I*-Ac: ¹H NMR (250 MHz, CDCl₃): $\delta = 1.97$ (s, 3 H, Ac), 3.20 (dd, ²J = 9.3 Hz, 1 H, SCHCH), 3.31 (dd, ²J = 16.0 Hz, ³J = 7.6 Hz, 1 H, SCHCH), 3.45 (dd, ³ $J_{2,1} = 9.0$ Hz, ³ $J_{2,3} = 9.0$ Hz, 1 H, 2-H), 3.48–3.53 (m, 1 H, 5-H), 3.60 (dd, $J_{4,5} = J_{4,3} = 8.7$ Hz, 1 H, 4-H), 3.63–3.75 (m, 3 H, 3-H, 2 6-H), 3.91 (ddd, ³J = 9.0 Hz, 7.7 Hz, 5.5 Hz, 1 H, SCH), 4.50–4.94 (m, 9 H, 1-H, 4 PhCH₂), 6.21 (d, ³J = 5.5 Hz, 1 H, AcOCH), 7.17–7.44 (m, 24 H, C₆H₄, 4 Ph).

D,L-erythro-1-Ethoxy-2-(2,3,4,6-tetra-*O*-benzyl-1-deoxy-β-D-glucopyranosylthio)indane (13a): To compound 12h,I (148 mg, 0.21 mmol) and 15-crown-5 (43 μL, 0.21 mmol) in DMF (5 mL) were added NaH (5 mg, 0.21 mmol) and ethyl iodide (0.5 mL, 6.2 mmol). At the end of the reaction, the solution was diluted with Et₂O and washed with water. The organic layer was concentrated in vacuo. Chromatography (toluene/ethyl acetate, 14:1) yielded 13a (130 mg, 86%) as colorless solid. – TLC (toluene/ethyl acetate, 9:1): $R_{\rm f} = 0.57$. – ¹H NMR (250 MHz, CDCl₃): $\delta = 1.20$, 1.21 (2 t, ${}^3J = 7.0$ Hz, 3 H, CH₃), 3.24–3.76 (m, 10 H, CH₂Me, CH₂CS, 2-H, 3-H, 4-H, 5-H, 2 6-H), 3.83 (mc, 0.5 H, SCH), 3.96 (mc, 0.5 H, SCH), 4.50–4.99 (m, 10 H, 1-H, 9 PhCH), 7.12–7.41 (m, 24 H, C₆H₄, 4 Ph). – C₄₅H₄₈O₆S (716.9): calcd. C 75.39, H 6.75; found C 74.88, H 6.69.

D,L-*erythro*-1-Hydroxy-2-(2,3,4,6-tetra-*O*-benzyl-β-D-glucopyranosylthio)-1′,2′,3′,4-tetrahydronaphthalene (15): To compound $1^{[18]}$ (1.0 g, 1.8 mmol) and 15-crown-5 (0.36 mmol) in DMF (10 mL) was added NaH (43 mg, 1.8 mmol). After 10 min, $14^{[26]}$ (0.82 g, 3.6 mmol) was added. After 7 h, the solution was diluted with Et₂O and washed with water. The organic layer was concentrated in vacuo. Chromatography (toluene/ethyl acetate, 20:1) yielded 15 (0.70 g, 55%) as a colorless oil. – TLC (toluene/ethyl acetate, 20:1): $R_f = 0.36$. – 1 H NMR (250 MHz, CDCl₃): $\delta = 2.00-2.35$ (m, 2 H, 2 3′-H), 2.73 – 3.05 (m, 2 H, 2 4′-H), 3.36 – 3.75 (m, 8 H, 2-H, 3-H, 4-H, 5-H, 2 6-H, 2′-H, OH), 4.47 – 4.96 (m, 10 H, 1-H, 1′-H, 8 PhCH), 708 – 7.40 (m, 24 H, C₆H₄, 4 Ph). – C₄₄H₄₆O₆S (702.9): calcd. C 75.18, H 6.60; found C 74.87, H 6.66.

D,L-erythro-1-Ethoxy-2-(-2,3,4,6-tetra-*O*-benzyl-1-deoxy-β-D-glucopyranosylthio)-1',2',3',4-tetrahydronaphthalene (16a): To compound 15 (0.70 g, 1.0 mmol) and 15-crown-5 (0.20 mL, 1.0 mmol) in dioxane (10 mL) and DMF (2 mL) was added NaH (24 mg, 1.0 mmol). After 10 min, ethyl iodide (0.16 mL, 2.0 mmol) was added. After 4 h, the solution was diluted with ethyl acetate and washed with water. The organic layer was concentrated in vacuo. Chromatography (toluene/ethyl acetate, 20:1) yielded 16a (0.54 g, 74%) as a colorless oil. – TLC (toluene/ethyl acetate, 9:1): R_f = 0.51. – ¹H NMR (250 MHz, CDCl₃): δ 1.20, 1.21 (2 s, 3 H, CH₃), 2.04–2.44 (m, 2 H, 2 3'-H), 2.70–2.83 (m, 1 H, 4'-H), 2.98–3.09 (m, 1 H, 4'-H), 3.42–3.77 (m, 9 H, 2-H, 3-H, 4-H, 5-H, 2 6 H, 2'-H, CH₂Me), 4.47–4.97 (m, 10 H, 1-H, 1'-H, 8 PhCH), 7.12–7.39 (m, 24 H, C₆H₄, 4 Ph). – C₄₆H₅₀O₆S (731.0): calcd. C 75.59, H 6.89; found C 75.40, H 7.03.

Ethyl 2,3,4,6-Tetra-O-(3-methylbenzyl)- α/β -D-glucopyranoside (19α α/β): To compound 20 α/β ^[13] (600 mg, 0.809 mmol, α/β = 4:1) and EtOH (94 μL, 1.62 mmol) in CH₂Cl₂ (1.0 mL) at -78 °C was added trimethylsilyl triflate (0.2 м in CH₂Cl₂, 120 μL, 0.02 mmol). The reaction mixture was allowed to reach 0 °C over 1.5 h. After addition of NEt₃ and coevaporation with toluene, chromatography (at first, petroleum ether/ethyl acetate, 8:1) yielded 19α β (434 mg, 86%) as a colorless syrup. A change to petroleum ether/ethyl acetate, 6:1 gave 19α α (29 mg, 6%) as a colorless syrup.

Compound 19aα: TLC (petroleum ether/ethyl acetate, 4:1): $R_{\rm f} = 0.46$. $- [α]_{\rm D} = 15.5$ (c = 1.0, CHCl₃). $- {}^{1}{\rm H}$ NMR (250 MHz, CDCl₃): δ = 1.24 (t, 3 H, CHCH₃), 2.29–2.32 (m, 12 H, 4 PhCH₃), 3.47–3.79 (m, 7 H, OCH₂Me, 2-, 4-, 5-, 6-, 6'-H), 3.98 (dd, 1 H, 3-H), 4.39–4.44 (m, 2 H, 2 MePhCH), 4.57–4.64 (m, 2 H, 2 MePhCH), 4.74–4.82 (m, 4 H, 3 MePhCH, 1-H), 4.97 (d, ${}^{3}{\it J} = 10.7$ Hz, 1 H, MePhCH), 6.93 (m, 2 H, 2 MePh-H), 7.05–7.26 (m, 14 H, 14 MePhH).

Compound 19aβ: TLC (petroleum ether/ethyl acetate, 4:1): $R_{\rm f} = 0.50$. $- [\alpha]_{\rm D} = 8.0$ (c = 1.0, CHCl₃). $- {}^{\rm 1}{\rm H}$ NMR (250 MHz,

CDCl₃): $\delta = 1.30$ (t, 3 H, OCCH₃), 2.30 (m, 12 H, 4 PhCH₃), 3.41–3.78 (m, 7 H, CHMe, 2-, 3-, 4-, 5-, 6-, 6'-H), 4.05 (m, 1 H, CHMe), 4.40 (d, ${}^{3}J = 7.8$ Hz, 1 H, 1-H), 4.46–4.95 (m, 8 H, 4 MePhCH₂), 6.95 (m, 2 H, 2 MePhH), 7.09–7.24 (m, 14 H, 14 MePhH). – $C_{40}H_{48}O_6$ (624.8): calcd. C 76.89, H 7.74; found C 77.13, H 7.75.

2-Bromobenzyl 2,3,4,6-Tetra-*O*-benzyl-α/β-D-glucopyranoside (19cα/β): These compounds were prepared following the general procedures for Table 3 and 4.

S-[2,3,4,6-Tetra-O-(3-methylbenzyl)-β-D-glucopyranosyl] Thioacetate (21): To compound 18^[13] (500 mg, 0.84 mmol) in dist. SOCl₂ (1.6 mL, 22 mmol) was added DMF (8 µL, 0.10 mmol), and the mixture was warmed to 50 °C for 30 min. The reaction mixture was concentrated in vacuo and the resulting syrup was dried under high vacuum for 1 h. The residue was dissolved in DMF (1.5 mL) and potassium thioacetate (380 mg, 3.3 mmol) was added. After 2 d at room temp., the black reaction mixture was processed by the usual workup. After the first chromatography (petroleum ether/ ethyl acetate, 6:1), a second elution [first toluene/ethyl acetate, 40:1, then (giving the compound) with toluene/ethyl acetate, 20:1] yielded 21 (464 mg, 84%) as a slightly red solid. – M.p. 52 °C (Et₂O/petroleum ether at -18 °C). - TLC (toluene/ethyl acetate, 10:1): $R_{\rm f} =$ $0.58. - [\alpha]_D = 25.3$ (c = 1.0, CHCl₃). $- {}^{1}H$ NMR (250 MHz, CDCl₃): $\delta = 2.29-2.31$ (m, 12 H, 4 PhCH₃), 2.37 (s, 3 H, SCOCH₃), 3.49-3.74 (m, 6 H, 2-H, 3-H, 4-H, 5-H, 6-H, 6'-H), 4.43 (d, ${}^{2}J = 12.0 \text{ Hz}$, 1 H, MePhCH), 4.50 (d, ${}^{2}J = 10.6 \text{ Hz}$, 1 H, MePhCH), 4.60 (d, ${}^{2}J = 12.0 \text{ Hz}$, 1 H, MePhCH), 4.69–4.84 (m, 5 H, 5 MePhCH), 5.16 (d, ${}^{3}J = 10.1$ Hz, 1 H, 1-H), 6.94 (m, 2 H, 2 MePhH), 7.09-7.25 (m, 14 H, 14 MePhH). $-C_{40}H_{46}O_6S$ (654.9): calcd. C 73.36, H 7.08; found C 73.51, H 7.07.

D,L-erythro-1-Hydroxy-2-[2,3,4,6-tetra-O-(3-methylbenzyl)-β-Dglucopyranosylthiolindane (22hll): Compound 21 (3.00 g, 4.58 mmol) was suspended in MeOH (15 mL), and NaOMe (1 m in MeOH, 4.7 mL, 4.7 mmol) was added at room temp. After 40 min, MeOH was removed under vacuum and the resulting solid was dried under high vacuum for 45 min. The residue was dissolved under exclusion of O2 in DMF (8 mL) at room temp. trans-2-Bromo-1-hydroxyindane (1.45 g, 6.80 mmol) and 15-crown-5 (0.91 mL, 4.6 mmol) were added and the solution was stirred for 2 h. Addition of sat. NaHCO₃ solution was followed by the usual workup. Chromatography (petroleum ether/ethyl acetate, 6:1, then petroleum ether/ethyl acetate, 3.5:1) yielded 22hll (2.96 g, 87%, h/ l = 1:1) as a colorless oil. The solvent system toluene/ethyl acetate, 12:1 allowed partial/total separation of 22h and 22l by chromatography/middle-pressure chromatography, although 221 still contained 5% of a by-product only separable in the next step.

Compound 221: TLC (toluene/ethyl acetate, 10:1): $R_{\rm f} = 0.38$.

Compound 22h: M.p. 88 °C (Et₂O/petroleum ether). – TLC (toluene/ethyl acetate, 10:1): $R_{\rm f} = 0.44$. – [α]_D = 16.8 (c = 1.0, CHCl₃). – ¹H NMR (250 MHz, CDCl₃): δ = 2.31 (2 s, 6 H, 2 PhCH₃), 2.33 (2 s, 6 H, 2 PhCH₃), 3.03 (dd, ²J = 15.8 Hz, ³J = 8.9 Hz, 1 H, indane-3-H), 3.22 (dd, ²J = 15.8 Hz, ³J = 7.6 Hz, 1 H, indane-3'-H), 3.41–3.77 (m, 8 H, OH, indane-2-H, 2-H, 3-H, 4-H, 5-H, 6-H, 6'-H), 4.50–4.61 (m, 4 H, 3 MePhCH, 1-H), 4.73–4.92 (m, 5 H, 5 MePhCH), 5.04 (d, 1 H, indane-1-H), 7.00 (m, 2 H, 2 MePhH), 7.12–7.34 (m, 18 H, 14 MePhH, 4 indane-H). – C₄₇H₅₂O₆S (745.0): calcd. C 75.77, H 7.04; found C 75.54, H 7.07.

D,L-erythro-1-(2-Bromobenzyloxy)-2-[2,3,4,6-tetra-*O*-(3-methylbenzyl)-β-D-glucopyranosylthiolindane (23ch): To compound 22h (525 mg, 0.70 mmol) in DMF (2.0 mL) were added 2-bromob-

enzyl bromide (300 mg, 1.20 mmol) and NaH (30 mg, 1.25 mmol) at room temp. under exclusion of light. To quicken the reaction, 15-crown-5 (140 µL, 0.71 mmol) was added after 2 h. After a further 4 h, MeOH and then sat. NaHCO3 solution were added, followed by the usual workup. Chromatography yielded 23ch (540 mg, 84%) as a colorless solid. - M.p. 68 °C (Et₂O/petroleum ether). -TLC (toluene/ethyl acetate, 10:1): $R_f = 0.66$. $- [\alpha]_D = 2.0$ (c = 1.0, CHCl₃). - ¹H NMR (600 MHz, CDCl₃): $\delta = 2.26-2.31$ (4 s, 12 H, 4 PhCH₃), 3.30 (dd, ${}^{2}J = 15.7$ Hz, ${}^{3}J = 7.3$ Hz, 1 H, indane-3-H), 3.35 (dd, ${}^{2}J = 15.7$ Hz, ${}^{3}J = 7.3$ Hz, 1 H, indane-3'-H), 3.47 (5-H), 3.50 (2-H), 3.60 (4-H), 3.66 (6-H), 3.69 (3-H), 3.74 (6'-H), 4.00 (indane-2-H), 4.47 (d, ${}^{2}J = 11.9 \text{ Hz}$, 1 H, MePhCH), 4.53-4.55 (m, 2 H, 2 MePhCH), 4.62 (d, $^{3}J = 9.7$ Hz, 1 H, 1-H), 4.70-4.83 (m, 5 H, 3 MePhCH, BrPhCH₂), 4.90-4.92 (m, 2 H, 2 MePhCH), 4.96 (m, 1 H, indane-1-H), 7.00 (m, 2 H, 2 MePhH), 7.06-7.28 (m, 19 H, 14 MePhH, 4 indane-H, BrPhH), 7.37 (m, 1 H, BrPhH), 7.48 (m, 1 H, BrPhH), 7.65 (m, 1 H, BrPhH). - ¹³C NMR (150.9 MHz, CDCl₃, selected data): $\delta = 39.3$ (indane-3-C), 48.6 (indane-2-C), 69.4 (6-C), 70.1 (BrPhC), 78.0 (4-C), 79.1 (5-C), 82.0 (2-C), 83.4 (indane-1-C), 84.4 (1-C), 86.7 (3-C). C₅₄H₅₇BrO₆S (914.01): calcd. C 70.96, H 6.29; found C 70.61, H

D,L-erythro-1-(2-Bromobenzyloxy)-2-[2,3,4,6-tetra-O-(3-methylbenzyl)-β-D-glucopyranosylthiolindane (23cl): To compound 22l (425 mg, 0.57 mmol) in DMF (2.0 mL) at room temp. were added 2-bromobenzyl bromide (185 mg, 0.74 mmol) and NaH (20 mg, 0.8 mmol). After 2 h, purification as described for 23ch yielded 23cl (420 mg, 81%) as a colorless solid. - M.p. 105 °C (ethyl acetate/ petroleum ether). – TLC (toluene/ethyl acetate, 10:1): $R_{\rm f} = 0.66$. $- [\alpha]_D = -13.2 (c = 1.0, CHCl_3). - {}^{1}H NMR (600 MHz, CDCl_3):$ $\delta = 2.25 - 2.30 \text{ (4 s, 12 H, 4 PhCH₃)}, 3.32 \text{ (dd, } ^2J = 15.8 \text{ Hz, } ^3J =$ 7.3 Hz, 1 H, indane-3-H), 3.40 (dd, ${}^{2}J = 15.8$ Hz, ${}^{3}J = 7.3$ Hz, 1 H, indane-3'-H), 3.49 (2-H), 3.50 (5-H), 3.57 (4-H), 3.65 (6-H), 3.68 (3-H), 3.74 (6'-H), 3.87 (m, 1 H, indane-2-H), 4.50-4.56 (m, 3 H, 3 MePhCH), 4.61 (d, ${}^{3}J = 9.7$ Hz, 1 H, 1-H), 4.68–4.75 (m, 3 H, BrPhCH₂, MePhCH), 4.79–4.82 (m, 2 H, 2 MePhCH), 4.88-4.94 (m, 3 H, 2 MePhCH, indane-1-H), 7.01 (m, 2 H, 2 MePhH), 7.06-7.26 (m, 19 H, 14 MePhH, 4 indane-H, BrPhH), 7.38 (m, 1 H, BrPhH), 7.48 (m, 1 H, BrPhH), 7.65 (m, 1 H, BrPhH). $- {}^{13}$ C NMR (150.9 MHz, CDCl₃, selected data): $\delta = 40.3$ (indane-3-C), 50.53 (indane-2-C), 69.4 (6-C), 69.8 (BrPhC), 78.0 (4-C), 79.0 (5-C), 82.2 (2-C), 85.9 (1-C), 86.6 (3-C). $-C_{54}H_{57}BrO_6S$ (914.0): calcd. C 70.96, H 6.29; found C 70.63, H 6.33.

9-(3-Chloro-2-pyrazinyl)-9-hydroxyxanthene (25): To THF (70 mL) was added *n*BuLi (1.6 M in hexane, 7.0 mL, 11.2 mmol) at -10 $^{\circ}$ C. This was cooled down to -70 $^{\circ}$ C. After addition of 2,2,6,6tetramethylpiperidine (2.0 mL, 11.8 mmol), the reaction mixture was warmed up to 0 °C, and 30 min later cooled down to -70 °C. Chloropyrazine (24) (0.80 mL, 8.8 mmol) was added and the mixture was stirred for 2 h at -70 °C. 9-Xanthenone (1.96 g, 10 mmol) was added; the mixture was stirred for 2 h at -70 °C and quenched by addition of sat. NaHCO₃ solution. After warming, water was added and extraction performed with CH2Cl2. The organic phase was washed with water, 1 N hydrochloric acid, and sat. NaHCO₃ solution. Drying with MgSO₄ and concentration in vacuo yielded a solid, which was dissolved in CH₂Cl₂ (5 mL). Chromatography (petroleum ether/ethyl acetate, 2:1) yielded 25 (2.23 g, 82%) as a colorless solid. - M.p. 179 °C (CH₂Cl₂/Et₂O). - TLC (petroleum ether/ethyl acetate, 2:1): $R_f = 0.28$. – ¹H NMR (250 MHz, CDCl₃): $\delta = 6.93 - 7.05$ (m, 4 H, 4 xanthene-H), 7.14 (s, 1 H, OH), 7.23 (m, 2 H, 2 xanthene-H), 7.34 (m, 2 H, 2 xanthene-H), 8.43 (d, ${}^{2}J =$ 2.5 Hz, 1 H, pyrazine-H), 8.61 (d, ${}^{2}J = 2.5$ Hz, 1 H, pyrazine-H).

 $-C_{17}H_{11}CIN_2O_2$ (310.7): calcd. C 65.71, H 3.57, N 9.01; found C 65.69, H 3.52, N 8.98.

9-Hydroxy-9-[3-(2,3,4,6-tetra-O-benzyl-β-D-glucopyranosylthio)-2pyrazinyl|xanthene (26): To compound 1 (212 mg, 0.38 mmol), suspended in MeOH (2 mL) was added NaOMe (1 m in MeOH, 380 μL, 0.38 mmol). After 10 min of stirring at room temp., the now clear solution was concentrated under high vacuum and dried for 45 min. The solid residue was dissolved under exclusion of O₂ in DMF (1.5 mL). Compound 25 (118 mg, 0.380 mmol) and 15crown-5 (80 µL, 0.40 mmol) were added. After 24 h, the usual workup followed. Chromatography (petroleum ether/ethyl acetate, 2:1) yielded 26 (264 mg, 83%) as a colorless, solid foam. - TLC (petroleum ether/ethyl acetate, 2:1): $R_f = 0.27$. $- [\alpha]_D = -9.6$ (c =1.0, CHCl₃). - ¹H NMR (250 MHz, CDCl₃): $\delta = 3.31-3.61$ (m, 6 H, 2-H, 3-H, 4-H, 5-H, 6-H, 6'-H), 4.16 (d, ${}^{2}J = 10.5$ Hz, 1 H, PhCH), 4.30-4.49 (m, 4 H, 4 PhCH), 4.72 (m, 3 H, 3 PhCH), 5.50 $(d, {}^{3}J = 10.2 \text{ Hz}, 1 \text{ H}, 1 \text{-H}), 6.77 - 7.34 \text{ (m, 28 H, 4 Ph, 8 xanthene-}$ H), 7.39 (s, 1 H, OH), 8.36 (d, ${}^{2}J = 2.6$ Hz, 1 H, pyrazine-H), 8.40 $(d, {}^{2}J = 2.6 \text{ Hz}, 1 \text{ H}, \text{ pyrazine-H}). - C_{51}H_{46}N_{2}O_{7}S (831.0)$: calcd. C 73.71, H 5.58, N 3.37; found C 73.62, H 5.57, N 3.53.

9-Ethoxy-9-[3-(2,3,4,6-tetra-*O***-benzyl-β-D-glucopyranosylthio)-2-pyrazinyl|xanthene (27a):** To compound **26** (600 mg, 0.72 mmol) in EtOH (6 mL) and CH₂Cl₂ (2 mL) was added camphor-10-sulfonic acid (20 mg, 0.09 mmol) at room temp. After 3 h, enough NEt₃ to ensure basic conditions was added. After concentration in vacuo, chromatography (petroleum ether/ethyl acetate, 3:1, 1 vol-% NEt₃) yielded **27a** (560 mg, 91%) as a colorless, solid foam. – TLC (petroleum ether/ethyl acetate, 3:1): $R_{\rm f} = 0.27$. – [α]_D = 41.4 (c = 1.0, CHCl₃). – ¹H NMR (250 MHz, CDCl₃): δ = 1.20 (t, ³J = 7.0 Hz, 3 H, CH₃), 3.12 (q, ³J = 7.0 Hz, 2 H, CH₂Me), 3.59–3.80 (m, 6 H, 2-H, 3-H, 4-H, 5-H, 6-H, 6'-H), 4.43 (d, ²J = 12.1 Hz, 1 H, PhCH), 4.55 (m, 2 H, 2 PhCH), 4.67–4.93 (m, 5 H, 5 PhCH), 5.65 (d, ³J = 10.2 Hz, 1 H, 1-H), 6.88–7.39 (m, 28 H, 4 Ph, 8 xanthene-H), 8.13 (s, 2 H, 2 pyrazine-H). – C₅₃H₅₀N₂O₇S (859.1): calcd. C 74.10, H 5.87, N 3.26; found C 73.74, H 5.86, N 3.35.

9-(Methyl 2,3,4-tri-O-benzyl-α-D-glucopyranosid-6-yl)-9-[3-(2,3,4,6tetra-O-benzyl-β-D-glucopyranosylthio)-2-pyrazinyl|xanthene (27b): To methyl 2,3,4-tri-O-benzyl-α-D-glucopyranoside (205 mg, 0.44 mmol), compound 26 (350 mg, 0.42 mmol), and molecular sieves (4 Å, 5 g) in CH₂Cl₂ (10 mL) at room temp. was added camphor-10-sulfonic acid (4 \times 25 mg, 4 \times 0.108 mmol) in four portions at hourly intervals. After 19 h, NEt₃ was added to the then neutral reaction mixture. The solution was separated from the molecular sieves, which were washed with ethyl acetate. After filtration, the combined extracts were concentrated and co-evaporated with toluene. Chromatography (toluene/ethyl acetate, 20:1, 1 vol-% NEt₃) yielded 27b (439 mg, 82%) as a colorless, solid foam. - TLC (toluene/ethyl acetate, 10:1): $R_f = 0.24$. $- [\alpha]_D = 34.5$ (c = 1.0, CHCl₃ + 1 vol-\% pyridine). - \frac{1}{4} NMR (250 MHz, CDCl₃ + 1 vol-\% [D₅]pyridine): $\delta = 3.21$ (dd, ${}^{2}J = 10.0$ Hz, ${}^{3}J = 5.7$ Hz, 1 H, 6b-H), 3.33-3.79 (m, 13 H, OMe, 2a-H, 2b-H, 3a-H, 4a-H, 4b-H, 5a-H, 5b-H, 6a-H, 6'a-H, 6'b-H), 3.94 (dd, 1 H, 3b-H), 4.22 (d, ${}^{2}J =$ 10.9 Hz, 1 H, PhCH), 4.42 (d, ${}^{2}J = 12.0$ Hz, 1 H, PhCH), 4.53 (d, $^{2}J = 12.0 \text{ Hz}, 1 \text{ H}, \text{ PhCH}, 4.57 - 4.86 (m, 11 \text{ H}, 10 \text{ PhCH}, 1b-H),}$ 4.93 (d, ${}^{2}J = 10.7$ Hz, 1 H, PhCH), 5.67 (d, ${}^{3}J = 10.3$ Hz, 1 H, 1a-H), 6.67 (m, 1 H, xanthene-H), 6.82 (m, 2 H, 2 xanthene-H), 6.96 (m, 1 H, xanthene-H), 7.08-7.40 (m, 39 H, 7 Ph, 4 xanthene-H), 8.09 (d, ${}^{3}J = 2.3$ Hz, 1 H, pyrazine-H), 8.13 (d, ${}^{3}J = 2.3$ Hz, 1 H, pyrazine-H). - C₇₉H₇₆N₂O₁₃S (1277.5): calcd. C 74.27, H 6.00, N 2.19; found C 73.96, H 6.09, N 2.49.

2-(2,3,4,6-Tetra-O-benzyl-β-D-glucopyranosylthio)pyrazine (28): To compound 1 (285 mg, 0.51 mmol) in DMF (1.5 mL) at 0 °C was

added NaH (14 mg, 0.58 mmol). Chloropyrazine (24) (73 µL, 0.82 mmol) was added after 10 min, the ice-bath was removed, and the reaction was stirred for 2 h at room temp. Addition of sat. NaHCO₃ solution was followed by the usual workup. Chromatography (toluene/ethyl acetate, 8:1) yielded 28 (282 mg, 87%) as a colorless solid. — TLC (toluene/ethyl acetate, 8:1): $R_{\rm f}=0.23$. — [α]_D = 15.0 (c=1.0, CHCl₃). — ¹H NMR (250 MHz, CDCl₃): $\delta=3.58-3.83$ (m, 6 H, 2-H 3-H, 4-H, 5-H, 6-H, 6'-H), 4.46 (d, $^2J=12.1$ Hz, 1 H, PhCH), 4.55—4.60 (m, 2 H, 2 PhCH), 4.77—4.95 (m, 5 H, 5 PhCH), 5.46 (d, $^2J=9.7$ Hz, 1 H, 1-H), 7.14—7.33 (m, 20 H, 4 Ph), 8.26 (m, 1 H, pyrazine-H), 8.36 (m, 1 H, pyrazine-H), 8.55 (m, 1 H, pyrazine-H). — $C_{38}H_{38}N_2O_5S$ (634.8): calcd. C 71.90, H 6.03, N 4.41; found C 71.55, H 6.09, N 4.42.

9-Hydroxy-9-{3-[2,3,4,6-tetra-O-(3-methylbenzyl)-β-D-glucopyranosylthio|-2-pyrazinyl|xanthene (29): To compound 21 (633 mg, 0.97 mmol), suspended in MeOH (1.5 mL), was added NaOMe (1 м in MeOH, 1 mL, 1 mmol). After 20 min at room temp., the clear solution was concentrated under high vacuum and dried for 45 min. The residue was dissolved in DMF (2 mL) under exclusion of O₂. Compound **25** (300 mg, 0.97 mmol) and 15-crown-5 (192 μL, 0.97 mmol) were added and the mixture was stirred for 13 h. The usual workup followed. Chromatography (petroleum ether/ ethyl acetate, 3:1) yielded 29 (826 mg, 96%) as a colorless syrup. -TLC (petroleum ether/ethyl acetate, 3:1): $R_{\rm f} = 0.28. - [\alpha]_{\rm D} = -$ 6.5 (c = 1.0, CHCl₃). – ¹H NMR (250 MHz, CDCl₃): $\delta =$ 2.24-2.28 (m, 12 H, 4 PhCH₃), 3.33 (dd, ${}^{3}J = 10.1$ Hz, ${}^{3}J =$ 9.8 Hz, 1 H, 2-H), 3.44-3.57 (m, 5 H, 3-H, 4-H, 5-H, 6-H, 6'-H), $4.12 \text{ (d, }^2J = 10.4 \text{ Hz, } 1 \text{ H, MePhCH)}, 4.27 \text{ (d, }^2J = 12.0 \text{ Hz, } 1 \text{ H,}$ MePhCH), 4.29 (d, ${}^{2}J = 10.4$ Hz, 1 H, MePhCH), 4.40 (d, ${}^{2}J =$ 12.0 Hz, 1 H, MePhCH), 4.43 (d, ${}^{2}J = 10.6$ Hz, 1 H, MePhCH), 4.63-4.73 (m, 3 H, 3 MePhCH), 5.50 (d, $^{3}J = 10.1$ Hz, 1 H, 1-H), 6.71-7.33 (m, 24 H, 16 MePhH, 8 xanthene-H), 7.38 (s, 1 H, OH), 8.35 (d, ${}^{3}J$ = 2.5 Hz, 1 H, pyrazine-H), 8.39 (d, ${}^{3}J$ = 2.5 Hz, 1 H, pyrazine-H). - C₅₅H₅₄N₂O₇S (887.1): calcd. C 74.47, H 6.14, N 3.16; found C 74.81, H 6.21, N 3.28.

9-(2-Bromobenzyloxy)-9-{3-[2,3,4,6-tetra-O-(3-methylbenzyl)-β-Dglucopyranosylthio]-2-pyrazinyl}xanthene (30c): To compound 29 (420 mg, 0.47 mmol), 2-bromobenzyl alcohol (89 mg, 0.47 mmol), and molecular sieves (4 Å, 3.0 g) in CH₂Cl₂ (5 mL) was added camphor-10-sulfonic acid (2 \times 55 mg, 2 \times 0.24 mmol), in two portions, separated by 2 h. After 2 h, the then neutral reaction mixture was processed as described for 27b. Chromatography (petroleum ether/ ethyl acetate, 5:1, 1 vol-% NEt₃) yielded 448 mg (90%) of 30c (448 mg, 90%) as a colorless syrup. - TLC (petroleum ether/ethyl acetate, 4:1): $R_f = 0.42$. $- [\alpha]_D = 11.1$ (c = 1.0, CHCl₃ + 1 vol-% pyridine). $- {}^{1}$ H NMR (250 MHz, CDCl₃ + 1 vol-% [D₅]pyridine): $\delta = 2.24$ (s, 3 H, PhCH₃), 2.27–2.29 (m, 9 H, 3 PhCH₃), 3.47–3.76 (m, 6 H, 2-H, 3-H, 4-H, 5-H, 6-H, 6'-H), 4.23 (dd, 2 H, BrPhCH₂), 4.38 (d, ${}^{2}J = 12.0 \text{ Hz}$, 1 H, MePhCH), 4.44 (d, ${}^{2}J = 10.4 \text{ Hz}$, 1 H, MePhCH), 4.50 (d, ${}^{2}J = 10.4$ Hz, 1 H, MePhCH), 4.52 (d, ${}^{2}J =$ 12.0 Hz, 1 H, MePhCH), 4.74-4.86 (m, 4 H, 2 MePhCH₂), 5.65 $(d, {}^{3}J = 10.3 \text{ Hz}, 1 \text{ H}, 1\text{-H}), 6.90-7.54 \text{ (m, 26 H, 16 MePhH, 8)}$ xanthene-H), 7.62 (m, 2 H, 2 BrPhH), 8.09 (d, ${}^{3}J = 2.3$ Hz, 1 H, pyrazine-H), 8.13 (d, ${}^{3}J = 2.3 \text{ Hz}$, 1 H, pyrazine-H).

2,7-Di-*tert***-butyl-4,5-bis(hydroxymethyl)-9,9-dimethylxanthene (32):** To compound **31** (3.36 g, 8.19 mmol) in THF (50 mL), in a reaction flask with a 12-inch Vigreux column, was added borane—dimethyl sulfide (10 M in THF, 3.3 mL, 33 mmol) at room temp. The reaction mixture was heated to reflux and the dimethyl sulfide was distilled off. After 4 h, the reaction mixture was cooled and water (30 mL) was added carefully. The mixture was extracted with Et₂O (2 \times 200 mL). The combined organic phase was washed twice with sat.

NaHCO₃ solution, then with water and brine. After drying with MgSO₄, concentration in vacuo gave a solid, which was dissolved in refluxing ethyl acetate (15 mL). After cooling and addition of petroleum ether (40 mL), **32** (2.76 g, 88%) crystallized as colorless crystals. Chromatography (petroleum ether/ethyl acetate, 3:2) of the concentrated mother liquor yielded further **32** (0.28 g, 9%). – M.p. 196° (ethyl acetate). – TLC (petroleum ether/ethyl acetate, 1:1): $R_f = 0.58$. – ¹H NMR (250 MHz, CDCl₃): $\delta = 1.33$ (s, 18 H, 2 tBu), 1.65 (s, 6 H, 2 Me), 2.38 (br. s, 1 H, 2 OH), 4.80 (s, 4 H, CH₂O), 7.18 (d, ⁴J = 2.3 Hz, 2 H, 2 xanthene-H), 7.40 (d, ⁴J = 2.3 Hz, 2 H, 2 xanthene-H). – EI-MS (70 eV); mlz (%): 382 (9) [M]⁺, 367 (100) [M – CH₃]⁺. – C₂₅H₃₄O₃ (382.5): calcd. C 78.49, H 8.96; found C 78.17, H 9.20.

2,7-Di-*tert***-butyl-4,5-bis(chloromethyl)-9,9-dimethylxanthene** (33): To dist. SOCl₂ (2.1 mL, 29 mmol) in CH₂Cl₂ (17 mL) was added 32 (1.84 g, 4.82 mmol) in CH₂Cl₂ (17 mL) over a period of 30 min at room temp. After 2 h, the reaction mixture was concentrated in vacuo. The usual workup, without chromatography, yielded 33 (1.86 g, 92%) as a colorless solid. – M.p. 202 °C (Et₂O/petroleum ether). – TLC (petroleum ether/ethyl acetate, 10:1): $R_{\rm f} = 0.71$. – ¹H NMR (250 MHz, CDCl₃): $\delta = 1.34$ (s, 18 H, 2 *t*Bu), 1.64 (s, 6 H, 2 Me), 4.87 (s, 4 H, 2 CH₂Cl), 7.28 (d, ⁴*J* = 2.3 Hz, 2 H, 2 xanthene-H), 7.41 (d, ⁴*J* = 2.3 Hz, 2 H, 2 xanthene-H). – C₂₅H₃₂Cl₂O (419.4): calcd. C 71.59, H 7.69; found C 71.80, H 7.65.

2,7-Di-tert-butyl-4-ethoxymethyl-5-hydroxymethyl-9,9-dimethylxanthene (34a): To compound 32 (350 mg, 0.92 mmol) in CH₂Cl₂ (20 mL) at −20 °C were added NaH (60% in oil, 46 mg, 1.15 mmol), 15-crown-5 (200 μL, 1.00 mmol), and ethyl triflate (120 μL, 0.93 mmol). After warming to 14 °C over a period of 2 h, stirring was continued for another 1 h at 14 °C. The addition of sat. NH₄Cl solution was followed by extraction with CH₂Cl₂. The organic phase was washed with water and brine, dried with MgSO₄, and concentrated in vacuo. Chromatography (toluene/ethyl acetate, 15:1) yielded **34a** (340 mg, 91%) as a colorless solid. – M.p. 190 °C (Et₂O/petroleum ether). – TLC (toluene/ethyl acetate, 3:1): $R_{\rm f} = 0.58. - {}^{1}{\rm H} \ {\rm NMR} \ (250 \ {\rm MHz}, \ {\rm CDCl_3}): \delta = 1.21 \ (t, 3 \ {\rm H},$ CHCH₃), 1.33 (s, 9 H, tBu), 1.34 (s, 9 H, tBu), 1.64 (2 s, 6 H, xanthene-CH₃), 3.50 (q, 2 H, CH₂Me), 4.68 (s, 2 H, CH₂O), 4.77 (s, 2 H, CH₂O), 7.16 (m, 2 H, xanthene-H), 7.37 (d, 1 H, xanthene-H), 7.41 (d, 1 H, xanthene-H). – EI-MS (70 eV); m/z (%): 410 (9) $[M]^+$, 395 (100) $[M - CH_3]^+$). $- C_{27}H_{38}O_3$ (410.7): calcd. C 78.98, H 9.33; found C 78.96, H 9.23.

2,7-Di-*tert***-butyl-4-chloromethyl-5-ethoxymethyl-9,9-dimethyl-xanthene (35a):** To compound **34a** (235 mg, 0.57 mmol) in CH₂Cl₂ (6 mL) at room temp. was added dist. SOCl₂ (125 μL, 1.7 mmol). After 2 h, the reaction mixture was concentrated in vacuo. Chromatography (petroleum ether/ethyl acetate, 40:1) yielded **35a** (218 mg, 89%) as a colorless syrup. – TLC (petroleum ether/ethyl acetate, 40:1): $R_f = 0.64$. – ¹H NMR (250 MHz, CDCl₃): $\delta = 1.31$ (t, ³J = 7.0 Hz, 3 H, OCCH₃), 1.33 (s, 9 H, tBu), 1.34 (s, 9 H, tBu), 1.64 (2 s, 6 H, 2 xanthene-CH₃), 3.69 (q, ³J = 7.0 Hz, 2 H, CH₂Me), 4.77 (s, 2 H, xanthene-CH₂), 4.79 (s, 2 H, xanthene-CH₂), 7.25 (d, 1 H, xanthene-H), 7.34 (d, 2 H, xanthene-H), 7.40 (d, ⁴J = 2.3 Hz, 1 H, xanthene-H). – EI-MS (70 eV); m/z (%): 428 (7) [M]⁺, 413 (100) [M – CH₃]⁺. – C₂₇H₃₇ClO₂ (429.0): calcd. C 75.59, H 8.69; found C 75.71, H 8.69.

2,7-Di-*tert*-butyl-4-chloromethyl-9,9-dimethyl-5-(methyl 2,3,4-tri-*O*-benzyl-α-D-glucopyranosid-6-yloxy)methylxanthene (35b): To methyl 2,3,4-tri-*O*-benzyl-α-D-glucopyranoside (890 mg, 1.92 mmol) in THF (20 mL) at room temp. were added NaH (60% in oil, 96 mg, 2.4 mmol), 15-crown-5 (0.80 mL, 4.0 mmol), compound 33

(810 mg, 1.93 mmol), and anhydrous NaI (288 mg, 1.92 mmol). After 2 d under exclusion of light, sat. NaHCO₃ solution was added and the usual workup followed. Chromatography (toluene/ethyl acetate, 24:1) gave as first fraction **35b** (678 mg, 41%) as a colorless syrup. – TLC (toluene/ethyl acetate, 10:1): $R_{\rm f} = 0.60$. – $[\alpha]_{\rm D} = 19.0$ (c = 1.0, CHCl₃). – ¹H NMR (250 MHz, CDCl₃): $\delta = 1.29$ (s, 9 H, tBu), 1.32 (s, 9 H, tBu), 1.54 (s, 3 H, xanthene-CH₃), 1.58 (s, 3 H, xanthene-CH₃), 3.38 (s, 3 H, OMe), 3.54–3.84 (m, 5 H, 2-H, 4-H, 5-H, 6-H, 6'-H), 3.96 (dd, 1 H, 3-H), 4.37 (d, ${}^{3}J = 10.6$ Hz, 1 H, PhCH), 4.63–5.00 (m, 10 H, 5 PhCH, 2 xanthene-CH₂, 1-H), 6.98–7.38 (m, 19 H, 3 Ph, 4 xanthene-H). – C₅₃H₆₃ClO₇ (847.5): calcd. C 75.11, H 7.49; found C 74.86, H 7.46.

2,7-Di-tert-butyl-4-chloromethyl-9,9-dimethyl-5-(methyl 2,3,6-tri-Obenzyl-\alpha-D-glucopyranosid-4-yloxy)methylxanthene methyl 2,3,6-tri-O-benzyl-α-D-glucopyranoside (1.11 g, 2.38 mmol) in THF (25 mL) at room temp. were added NaH (60% in oil, 170 mg, 4.3 mmol), 15-crown-5 (0.95 mL, 4.8 mmol), compound 33 (1.00 g, 2.38 mmol), and anhydrous NaI (360 mg, 2.40 mmol). The reaction was treated as described for 35b. The first fraction was 35d (1.29 g, 64%) as a colorless syrup. - TLC (toluene/ethyl acetate. 10:1): $R_{\rm f} = 0.59$. $- [\alpha]_{\rm D} = 57.0 \ (c = 1.0, \text{ CHCl}_3)$. $- {}^{1}\text{H} \ \text{NMR}$ (250 MHz, CDCl₃): $\delta = 1.30$ (s, 9 H, tBu), 1.33 (s, 9 H, tBu), 1.60 (s, 3 H, xanthene-CH₃), 1.63 (s, 3 H, xanthene-CH₃), 3.40 (s, 3 H, OMe), 3.56-3.70 (m, 4 H, 2-H, 4-H, 6-H, 6'-H), 3.82 (m, 1 H, 5-H), 4.03 (dd, 1 H, 3-H), 4.40 (s, 2 H, xanthene-CH₂), 4.54 (d, ${}^{3}J =$ 11.3 Hz, 1 H, PhCH), 4.62-4.99 (m, 7 H, 1-H, xanthene-CH₂, 4 PhCH), 5.12 (d, ${}^{3}J = 12.7 \text{ Hz}$, 1 H, PhCH), 7.11–7.39 (m, 19 H, 4 xanthene-H, 3 Ph). - C₅₃H₆₃ClO₂ (847.5): calcd. C 75.11, H 7.49; found C 75.04, H 7.56.

2,7-Di-tert-butyl-5-ethoxymethyl-9,9-dimethyl-4-(2,3,4,6-tetra-Obenzyl-β-D-glucopyranosylthio)methylxanthene (36a): To compound 1 (187 mg, 0.33 mmol) and compound 35a (142 mg, 0.33 mmol) in DMF (4 mL) at 0 °C were added NaH (17 mg, 0.7 mmol) and 15crown-5 (66 µL, 0.33 mmol). After 2 h, MeOH and then sat. NH₄Cl solution were added and followed by the usual workup. Chromatography (toluene/ethyl acetate, 20:1) yielded 36a (307 mg, 97%) as a colorless syrup. – TLC (petroleum ether/ethyl acetate, 4:1): $R_{\rm f}$ = $0.60. - [\alpha]_D = -83.7$ (c = 1.0, CHCl₃). $- {}^{1}$ H NMR (600 MHz, CDCl₃): $\delta = 1.24$ (t, 3 H, CHCH₃), 1.27 (s, 9 H, tBu), 1.35 (s, 9 H, tBu), 1.58 (s, 3 H, xanthene-CH₃), 1.65 (s, 3 H, xanthene-CH₃), 3.45 (5-H), 3.47 (2-H), 3.59 (3-H), 3.59 (4-H), 3.60 (m, 2 H, CH₂Me), 3.71 (6-H), 3.80 (6'-H), 4.11 (m, 2 H, CH₂S), 4.35 (d, $^{3}J = 9.5 \text{ Hz}, 1 \text{ H}, 1 \text{-H}, 4.52 - 4.88 (m, 10 \text{ H}, 4 \text{ PhCH}_{2}, \text{CH}_{2}\text{OEt}),$ 6.94 (m, 2 H, 2 PhH), 7.06-7.40 (m, 24 H, 4 Ph, 4 xanthene-H (3: 7.12, 6: 7.21)). - 13C NMR (150.9 MHz, CDCl₃, selected data): $\delta = 15.3 \text{ (OCCH}_3), 29.1 \text{ (CH}_2\text{S)}, 66.2 \text{ (OCH}_2\text{Me)}, 67.9 \text{ (CH}_2\text{OEt)},$ 69.2 (6-C), 78.1 (4-C), 79.0 (5-C), 81.8 (2-C), 82.7 (1-C), 86.6 (3-C). – INEPT: ${}^{1}J_{1-H,1-C} = 157 \text{ Hz.} - C_{61}H_{72}O_{7}S$ (949.3): calcd. C 77.18, H 7.64; found C 77.36, H 7.63.

2,7-Di-*tert*-butyl-9,9-dimethyl-5-(methyl 2,3,4-tri-*O*-benzyl-α-D-glucopyranosid-6-yloxy)methyl-4-(2,3,4,6-tetra-*O*-benzyl-β-D-glucopyranosylthio)methylxanthene (36b): To compound 1 (328 mg, 0.59 mmol) and compound 35b (500 mg, 0.59 mmol) in DMF (5 mL) at 0 °C were added NaH (24 mg, 1.0 mmol) and 15-crown-5 (117 μL, 0.59 mmol). Ice-cooling was removed after 1 h and the reaction was stirred for 20 h at room temp. The reaction was worked up by addition of sat. NaHCO₃ solution and water, followed by extraction with Et₂O. The organic phase was washed with water and brine, dried with MgSO₄, and concentrated in vacuo. Chromatography (toluene/ethyl acetate, 5:1) yielded 36b (640 mg, 79%) as a colorless syrup. – TLC (toluene/ethyl acetate, 10:1): R_f = 0.35. – [α]_D = - 65.6 (c = 1.0, CHCl₃). – ¹H NMR (600 MHz,

CDCl₃): $\delta = 1.27$ (s, 9 H, tBu), 1.30 (s, 9 H, tBu), 1.54 (s, 3 H, xanthene-CH₃), 1.63 (s, 3 H, xanthene-CH₃), 3.35 (s, 3 H, OMe), 3.43 (5a-H), 3.44 (2a-H), 3.55 (2b-H), 3.56 (3a-H), 3.59 (4a-H), 3.60 (4b-H), 3.69 (6b-H), 3.70 (5b-H), 3.71 (6a-H), 3.75 (6b'-H), 3.80 (6'a-H), 3.94 (3b-H), 4.09 (m, 2 H, CH_2S), 4.31 (d, 3J = 9.8 Hz, 1 H, 1a-H), 4.43-4.48 (m, 2 H, 2 PhCH), 4.55 (d, ${}^{2}J =$ 10.7 Hz, 1 H, PhCH), 4.61 (d, 1 H, 1b-H), 4.61-4.66 (m, 4 H, 4 PhCH), 4.71 (d, ${}^{2}J = 10.6$ Hz, 1 H, PhCH), 4.75–4.84 (m, 5 H, 5 PhCH), 4.86 (s, 2 H, xanthene-CH₂O), 4.94 (d, ${}^{2}J = 11.0 \text{ Hz}$, 1 H, PhCH), 6.89 (m, 2 H, 2 PhH), 7.05-7.41 (m, 37 H, 33 Ph-H, 4 xanthene-H (3: 7.10, 6: 7.34)). - 13C NMR (150.9 MHz, CDCl₃, selected data): $\delta = 29.2$ (CH₂S), 68.1 (xanthene-CH₂O), 68.7 (6b-C), 69.2 (6a-C), 70.2 (5b-C), 77.7 (4b-C), 78.0 (4a-C), 78.9 (5a-C), 79.9 (2b-C), 81.8 (2a-C), 82.1 (3b-C), 82.4 (1a-C), 86.6 (3a-C), 98.1 (1b-C). – INEPT: ${}^{1}J_{1a-H,1a-C} = 157 \text{ Hz}, {}^{1}J_{1b-H,1b-C} = 168 \text{ Hz}.$ – C₈₇H₉₈O₁₂S (1367.8): calcd. C 76.40, H 7.22; found C 76.39, H 7.27.

2,7-Di-tert-butyl-9,9-dimethyl-4-(2,3,4,6-tetra-O-benzyl-β-D-glucopyranosylthio)methyl-5-(methyl 2,3,6-tri-O-benzyl-α-D-glucopyranosid-4-yloxy)methylxanthene (36d): To compound 1 (668 mg, 1.20 mmol) and compound 35d (1.03 g, 1.22 mmol) in DMF (9 mL) at 0 °C were added NaH (36 mg, 1.5 mmol) and 15-crown-5 (240 μ L, 1.20 mmol). The reaction mixture was treated as described for 36b. Chromatography (toluene/ethyl acetate, 18:1) yielded 36d (1.53 g, 93%) as a colorless syrup. - TLC (toluene/ ethyl acetate, 10:1): $R_f = 0.57$. $- [\alpha]_D = -33.0$ (c = 1.0, CHCl₃). - ¹H NMR (600 MHz, CDCl₃): $\delta = 1.28$ (s, 9 H, tBu), 1.31 (s, 9 H, tBu), 1.54 (s, 3 H, xanthene-CH₃), 1.65 (s, 3 H, xanthene-CH₃), 3.36 (s, 3 H, OMe), 3.39 (5a-H), 3.41 (2a-H), 3.48 (6b-H), 3.55 (3a-H), 3.55 (2b-H), 3.58 (6'b-H), 3.60 (4b-H), 3.61 (4a-H), 3.69 (6a-H), 3.74 (6'a-H), 3.80 (5b-H), 3.96 (CH-S-1a-C), 4.01 (3b-H), 4.01 (CH-S-1a-C), 4.32 (1a-H), 4.32-4.38 (m, 2 H, 2 PhCH), 4.43 (d, $^{2}J = 9.8 \text{ Hz}$, 1 H, PhCH), 4.54–4.58 (m, 4 H, 2 PhCH₂), 4.58 (1b-H), 4.64 (d, ${}^{2}J = 12.0 \text{ Hz}$, 1 H, PhCH), 4.71–4.84 (m, 5 H, 5 PhCH), 4.93 (d, ${}^{2}J = 11.0 \text{ Hz}$, 1 H, PhCH), 5.03 (d, ${}^{2}J = 12.2 \text{ Hz}$, 1 H, CHO-4b-C), 5.08 (d, ${}^{2}J$ = 12.2 Hz, 1 H, CHO-4b-C), 6.86 (m, 2 H, 2 PhH), 7.00-7.26 (m, 37 H, 33 PhH, 4 xanthene-H (3: 7.11, 6: 7.30, 1: 7.31, 8: 7.35). – ¹³C NMR (150.9 MHz, CDCl₃, selected data): $\delta = 29.6$ (CH₂S), 55.1 (OMe), 69.2 (6a-C), 69.5 (6b-C), 69.9 (CO-4b-C), 70.2 (5b-C), 78.0 (4a-C), 78.5 (4b-C), 78.8 (5a-C), 80.2 (2b-C), 81.8 (2a-C), 82.2 (3b-C), 83.2 (1a-C), 86.7 (3a-C), 98.0 (1b-H). - C₈₇H₉₈O₁₂ (1367.8): calcd. C 76.40, H 7.22; found C 76.34,

2,7-Di-tert-butyl-9,9-dimethyl-5-(methyl 2,3,4-tri-O-benzyl-α-Dglucopyranosid-6-yloxy)methyl-4-(N-succinimidylthio)methylxanthene (37b): To compound 36b (114 mg, 0.08 mmol) and 2,6-di-tertbutyl-4-methylpyridine (85 mg, 0.41 mmol) in CH₂Cl₂ (3.0 mL) was added N-iodosuccinimide (93 mg, 0.41 mmol). After 16 h under exclusion of light, the addition of sat. Na₂S₂O₃ solution was followed by the usual workup. Chromatography (toluene/ethyl acetate, 5:1) yielded 22b (64 mg, 86%) as a colorless syrup. – TLC (toluene/ ethyl acetate, 5:1): $R_f = 0.23$. $- [\alpha]_D = 22.2$ (c = 1.0, CHCl₃). -¹H NMR (250 MHz, CDCl₃): $\delta = 1.29$ (s, 9 H, tBu), 1.30 (s, 9 H, tBu), 1.52 (s, 3 H, xanthene-CH₃), 1.56 (s, 3 H, xanthene-CH₃), 2.53 (s, 4 H, CH₂CH₂), 3.38 (s, 3 H, OMe), 3.55 (dd, 1 H, 6-H), 3.59-3.98 (m, 5 H, 2-H, 3-H, 4-H, 5-H, 6'-H), 4.20 (d, ${}^{2}J =$ 12.2 Hz, 1 H, CH-S), 4.26 (d, ${}^{2}J = 12.2$ Hz, 1 H, CHS), 4.40 (d, $^{2}J = 10.6 \text{ Hz}, 1 \text{ H}, \text{ PhCH}, 4.63 - 5.06 \text{ (m, 8 H, 5 PhCH, xanthene-$ CH₂, 1-H), 6.98-7.03 (m, 3 H, 3 PhCH), 7.16-7.37 (m, 16 H, 12 PhH, 4 xanthene-H). – FAB MS (positive mode, matrix: 3nitrobenzyl alcohol with NaI); m/z: 1115 (2) [{MNaI}Na]⁺, 964 (75), $[MNa]^+$, 478 (100). - $C_{57}H_{67}NO_9S$ (942.2): calcd. C 72.66, H 7.17, N 1.49; found C 72.51, H 7.20, N 2.00.

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- [1] L. M. Sinnott, Chem. Ber. 1990, 90, 1171-1202; G. Legler, Adv. Carbohydr. Chem. Biochem. 1990, 48, 319-384.
- R. R. Schmidt, H. Dietrich, Angew. Chem. 1991, 109, 1348-1349; Angew. Chem. Int. Ed. Engl. 1991, 30, 1328-1329;
 R. R. Schmidt, K. Frische, Bioorg. Med. Chem. Lett. 1993, 3, 1747-1750;
 F. Amann, C. Schaub, B. Müller, R. R. Schmidt, Chem. Eur. J. 1998, 4, 1106-1115;
 B. Müller, C. Schaub, R. R. Schmidt, Angew. Chem. 1998, 110, 3021-3024;
 Angew. Chem. Int. Ed. 1998, 37, 2893-2897.
- [3] F. Barresi, O. J. Hindsgaul, J. Am. Chem. Soc. 1991, 113, 9376–9377; Synlett 1992, 759–761; Can. J. Chem. 1994, 72, 1447–1465.
- [4] R. Preuss, K. -H. Jung, R. R. Schmidt, *Liebigs Ann. Chem.* 1992, 377–382.
- [5] G. Stork, G. J. Kim, J. Am. Chem. Soc. **1992**, 114, 1087–1088.
- [6] M. Bols, J. Chem. Soc., Chem. Commun. 1992, 913–914; Acta Chem. Scand. 1996, 50, 931–937.
- Y. Ito, T. Ogawa, Angew. Chem. 1994, 106, 1843–1845; Angew. Chem. Int. Ed. Engl. 1994, 33, 1765–1767; J. Am. Chem. Soc. 1997, 119, 5562–5566.
- [8] [8a] T. Ziegler, R. Lau, Tetrahedron Lett. 1995, 36, 1417-1420. [8b] R. Lau, G. Schüle, U. Schwaneberg, T. Ziegler, Liebigs Ann. 1995, 1745-1754. [8c] G. Schüle, T. Ziegler, Liebigs Ann. 1996, 1599-1607. [8d] T. Ziegler, G. Lemanski, Eur. J. Org. Chem. 1998, 163-170; Angew. Chem. 1998, 110, 3367-3369; Angew. Chem. Int. Ed. 1998, 37, 3129-3132.
- [9] [9a] S. Valverde, A. M. Gomez, A. Hernandez, B. Herradon, J. C. Lopez, J. Chem. Soc., Chem. Commun. 1995, 2005-2006. [9b]
 S. Valverde, A. M. Gomez, J. C. Lopez, B. Herradon, Tetrahedron Lett. 1996, 37, 1105-1108.
- [10] H. Yamada, K. Imamura, T. Takahashi, *Tetrahedron Let.* 1997, 38, 391–394.
- [11] [11a] R. R. Schmidt, M. Stumpp, Liebigs Ann. Chem. 1983, 1249-1259.
 [11b] R. R. Schmidt in Carbohydrates Synthetic Methods and Applications in Medicinal Chemistry (Eds.: H. Ogura, A. Hasegawa, T. Suami), Kodanasha Ltd., Tokyo, 1992, p. 68-88. [11c] M. E. Behrendt, R. R. Schmidt, Tetrahedron Lett. 1993, 34, 6733-6736.
- [12] T. Iimori, T. Shibazaki, S. Ikegami, *Tetrahedron Lett.* 1996, 37, 2267–2270.
- [13] G. Scheffler, R. R. Schmidt, Tetrahedron Lett. 1997, 38, 2943-2946; J. Org. Chem. 1999, 64, 1319-1325.

- [14] C. Mukai, T. Itoh, M. Hanaoka, Tetrahedron Lett. 1997, 38, 4595-4598.
- [15] U. Huchel, R. R. Schmidt, Tetrahedron Lett. 1998, 38, 7693-7694; M. Müller, U. Huchel, A. Geyer, R. R. Schmidt, J. Org. Chem. 1999, 64, 6190-6201.
- [16] M. E. Behrendt, Dissertation, Universität Konstanz, 1994.
- [17] G. Scheffler, Dissertation, Universität Konstanz, 1998.
- [18] S. A. Holick, L. Anderson, *Carbohydr. Res.* **1974**, *43*, 208–213.
- [19] F. N. Tebbe, G. W. Parshall, G. S. Reddy, J. Am. Chem. Soc. 1978, 100, 3611-3613; K. Clauss, H. Bestian, Justus Liebigs Ann. Chem. 1962, 654, 8-19.
- [20] F. Weygand, H., Ziemann, Justus Liebigs Ann. Chem. 1962, ((AUTHOR: Vol. number missing!)), 179-198.
- [21] R. R. Schmidt, M. Behrendt, A. Toepfer, Synlett 1990, 694-696.
- [22] Commercially available.
- ^[23] R. R. Schmidt, U. Moering, M. Reichrath, *Chem. Ber.* **1982**, 115, 39–49.
- [24] [24a] R. R. Schmidt, Angew. Chem. 1986, 98, 213–236; Angew. Chem. Int. Ed. Engl. 1986, 25, 212–235. [24b] K. Toshina, K. Tatsuta, Chem. Rev. 1993, 93, 1503–1531.
- [25] Y. D. Vankar, P. S. Vankar, M. Behrendt, R. R. Schmidt, *Tetrahedron* 1991, 47, 9985–9988; R. R. Schmidt, H. Gaden, H. Jatzke, *Tetrahedron Lett.* 1990, 31, 327–330.
- [26] D. R. Dalton, V. P. Dutta, D. C. Jones, J. Am. Chem. Soc. 1968, 90, 5498-5501
- [27] T. D. Perrine, C. P. J. Glaudemans, R. K. Ness, J. Kyle, H. G. Fletcher, Jr., J. Org. Chem. 1967, 32, 664-669.
- [28] R. R. Schmidt, in *Modern Methods in Carbohydrate Synthesis* (Eds.: S. H. Khan, R. A. O'Neill), Harwood Academic Publisher GmbH., Chur, Schweiz, 1996, p. 20–54.
- [29] G. Zemplén, Ber. Dtsch. Chem. Ges. 1927, 60, 1555-1564.
- [30] A. Turck, L. Mojovic, G. Quéguiner, Synthesis 1988, 881–884.
- [31] T. Mukaiyama, T. Nakatsuka, S. Shoda, *Chem. Lett.* **1979**, 487–490. [31a] R. B. Woodward et al., *J. Am. Chem. Soc.* **1981**, *103*, 3215–3217. [31b] S. Hanessian, A. Ugolini, D. Dube, P.J. Hodges, C. Andre, *J. Am. Chem. Soc.* **1986**, *108*, 2776–2778.
- [32] R. Eby, C. Schuerch, Carbohydr. Res. 1974, 34, 79-90; A. Lipták, J. Jodál, P. Nánási, Carbohydr. Res. 1975, 44, 1-11.
- [33] J. M. Küster, I. Dyong, Justus Liebigs Ann. Chem. 1975, 2179-2189; P. J. Garegg, H. Hultberg, S. Wallin, Carbohydr. Res. 1982, 108, 97-101.
- [34] R. R. Schmidt, J. Michel, J. Carbohydr. Res. 1985, 4, 141-169.
- [35] K. Mislow, J. Am. Chem. Soc. 1951, 73, 3954-3956; H. Takeuchi, K. Kitajiama, Y. Yamamoto, K. Mizuno, J. Chem. Soc., Perkin Trans. 2 1993, 199-203.

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